

UNIVERSIDADE ESTADUAL DE MONTES CLAROS

Lílian Mendes Borburema Cangussu

Potencial antineoplásico do ácido gálico em camundongos submetidos à exposição ao carcinógeno 4NQO e da nanopartícula AICIPc em células Scc9: duas novas propostas.

Montes Claros
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Área de Concentração:

Orientador: Prof. Dr. André Luiz Sena Guimarães

Coorientadora: Profa. Dra. Ludmilla Regina de Souza

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Nossa Senhora Aparecida,

São Padre Pio e

São Miguel Arcanjo,

Consagro.

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*A Tia Paula
Laís, Tia Dora e
Ivo,*

Dedico.

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Coordenador(a) Adjunto(a): Marise Fagundes



CANDIDATA: LILIAN MENDES BORBUREMA CANGUSSU

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BANCA (TITULARES)

PROF. DR. ANDRÉ LUIZ SENA GUIMARÃES - ORIENTADOR

PROF. DR. LUDMILLA REGINA DE SOUZA - COORIENTADORA

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APROVADO(A)

REPROVADO(A)

RESUMO

O câncer bucal causa morbidade e mortalidade significativas. Em todo o mundo, o carcinoma de células escamosas orais (CCEO) é o sexto câncer mais comum. As modalidades de tratamento atualmente disponíveis incluem cirurgia, radioterapia, quimioterapia e estratégias de prevenção, incluindo mudanças de estilo de vida, bem como quimioprevenção. A Terapia Fotodinâmica (TFD), por provocar a morte de células neoplásicas a partir da união da ação de um fotossensibilizante e de luz em comprimento de onda adequado, tem sido uma alternativa à quimioterapia e à radioterapia para o tratamento clínico do câncer devido aos seus efeitos locais que tornam o tratamento menos agressivo para o paciente. Este trabalho apresenta duas novas propostas para o tratamento de CCEO: um fotossensibilizante (AICIPc) como alternativa aos já existentes, de eficácias reduzidas e o Ácido Gálico, que por seus efeitos antioxidantes e anti-inflamatórios tem demonstrado possuir ação antineoplásica, o que o torna um agente promissor no tratamento de OSCC. A TFD foi realizada *in vitro* em células Scc9 nas quais foram realizados ensaios para avaliar o fenótipo celular após o tratamento com AICIPc. O Ácido Gálico foi administrado via gavagem em animais submetidos ao tratamento com o carcinógeno 4nqo. Os animais foram pesados diariamente para determinação da dose do AG introduzida por via oral até a morte dos animais. A TFD provocou morte celular considerável ($p < 0,05$) e o Ácido Gálico amenizou a gravidade das lesões instaladas nas línguas dos animais. Os resultados deste trabalho apresentam recursos que podem ser ponto de partida para novas alternativas para o tratamento do câncer bucal.

Palavras-chave: Terapia Fotodinâmica. 4nqo. Ação antineoplásica.

ABSTRACT

Oral cancer causes significant morbidity and mortality. Around the world, squamous cell carcinoma (OSCC) is the sixth most common cancer. Treatment modalities are available in surgery, radiotherapy, chemotherapy and prevention strategies, including lifestyle changes as well as chemoprevention. Photodynamic Therapy (PDT), due to the death of neoplastic cells, has been the target of photosensitization and light in the proper wave direction. It has been an alternative to chemotherapy and radiotherapy for the clinical treatment of cancer makes the treatment less aggressive for the patient. This work presented the new measures for the treatment of OSCC: a photosensitizer (AICIPc) as once existing, of reduced sessions and the Gallic Acid, which by its own antioxidants and anti-inflammatory effects has a more effective antineoplastic action, the which makes it a promising agent in the treatment of OSCC. PDT was performed in vitro on Scc9 cells in which assays were performed to evaluate the cellular phenotype after treatment with AICIPc. Gallic acid was administered gavage in animals after treatment with the carcinogen 4nqo. The animals were weighed daily for GA oral dosing until the animals died. A PDT caused cell death ($p < 0.05$) and Gallic acid attenuated lesions in the animals' tongues. The results of this work may be the starting point for new alternatives for the treatment of oral cancer.

Keywords: Photodynamic Therapy. 4nqo. Antineoplastic action.

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1 INTRODUÇÃO

De acordo com o Instituto Nacional do Câncer (1), a estimativa 2018-2019 para Carcinoma Epidermoide de Boca (CEB) é de 14.700 novos casos, sendo 11.200 homens e 3.500 mulheres. No mundo em 2018, os novos casos de câncer da cavidade oral alcançaram 354,864 e o número de óbitos 177,384 (2).

As Diretrizes da Política Nacional de Saúde Bucal (3) orientam que deve-se estabelecer parcerias para a prevenção, diagnóstico, tratamento e recuperação do câncer bucal com Universidades e outras organizações.

O Caderno de Atenção Básica do Ministério da Saúde (4) pressupõe que o tratamento cirúrgico e radioterápico deverá ser feito em nível de média e alta complexidade e recomenda que, a reabilitação deve permear a reposição de perdas estéticas e funcionais causadas pela doença.

Mesmo com procedimentos de reconstrução cirúrgica, uma das consequências após os tratamentos é a ocorrência de algum grau de desfiguração facial, promovendo graves defeitos estéticos e funcionais (5).

Em função da heterogeneidade das mudanças genéticas, o CEB é uma neoplasia de difícil tratamento. Tendo em vista as desvantagens apresentadas, faz-se necessário o desenvolvimento de novas alternativas que possibilitem maior segurança e efetividade terapêutica no processo de tratamento de indivíduos acometidos pela doença. Considerando as possibilidades terapêuticas atuais para o tratamento das neoplasias bucais, estudos têm demonstrado a utilização de terapias alternativas no controle da doença (6-10). A terapia fotodinâmica (TFD) é uma destas alternativas para o tratamento de lesões neoplásicas dos tecidos bucais (11, 12) e o Ácido Gálico diminuiu a resposta inflamatória em células neoplásicas (13) além de possuir uma variedade de atividades farmacológicas, como atividades antioxidantes e anticancerígenas em estudos pré-clínicos (14).

A Nanopartícula Alumínio Cloro Ftalocianina (AlClPc) é um fotossensibilizante quimicamente estável de segunda geração que possui propriedades fotofísicas que são adequadas para a realização de TFD como, altos rendimentos quânticos tripleto (formação de espécies reativas de oxigênio) (15). A (bio) química da TFD não pode ser separada das suas propriedades físicas, já que o efeito biológico/clínico é resultado das interações quantitativas das duas (16). Embora

AICIPc tenha alto peso molecular, sua natureza hidrofóbica facilita a interação com as camadas bilipídicas das células (15) tornando-a ideal para a realização da TFD.

A fonte de luz utilizada neste trabalho foi o diodo emissor de luz (LED), com comprimento de onda de 660nm. O aparelho permitiu a realização do trabalho com potência de 47.2 mW/cm² e frequência: 28.35 Jcm². A morte celular é dependente da densidade de energia. Na menor densidade de energia, a apoptose foi predominante, enquanto na densidade de energia mais alta, a necrose foi o mecanismo predominante de morte celular (17).

A TFD é um procedimento terapêutico clinicamente aprovado, minimamente invasivo, que pode exercer uma atividade citotóxica seletiva para células malignas (18). É um procedimento de dois estágios: após a administração de uma substância (fotossensibilizante) sensível à luz, o tumor é irradiado com uma luz de comprimento de onda apropriado. A luz pode ser entregue a praticamente qualquer órgão do corpo por meio de dispositivos flexíveis de fibra óptica (18). Existem três mecanismos principais pelos quais a TFD medeia a destruição do tumor que podem influenciar-se mutuamente (19): 1. As espécies reativas de oxigênio geradas pela TFD podem matar células tumorais diretamente; 2. a TFD danifica a vasculatura associada ao tumor, levando-o a morte; 3. A TFD pode ativar uma resposta imune contra células tumorais (20).

Dados recentes (21) dão conta de que a AICIPc em baixas concentrações interage com poucos sítios de DNA causando uma curvatura na estrutura da fita dupla, o que fornece um ambiente favorável para a intercalação de agregados do fotossensibilizante. A combinação de todos esses componentes é necessária para o controle do tumor a longo prazo (20).

Uma vez que a droga fotossensibilizante (DF) absorve a luz, atingindo um estado excitado, pode emitir fluorescência ou fosforescência, pode reagir com diferentes moléculas circundantes, gerando radicais - fotorreação tipo 1 - ou pode catalisar a conversão de oxigênio tripleto ($^3\text{O}_2$) na fotorreação de oxigênio singleto ($^1\text{O}_2$) - tipo 2 (22).

O Ácido Gálico (AG), conhecido como ácido 3, 4, 5-tri-hidroxibenzóico, está presente amplamente em diferentes espécies de plantas, inclusive na casca do fruto da *Caryocar brasiliense* com o equivalente a 209 gramas de Ácido Gálico por kg (23). Esta capacidade permitiu que o pré-tratamento de linfócitos com AG inibisse a apoptose induzida pelo estresse oxidativo por ação direta na eliminação de radicais

livres, antioxidante mais forte que o Trolox, um análogo da vitamina E solúvel em água (24).

Além disto, este composto provoca um efeito antiproliferativo nas células SCC uma vez que contribuiu para a inibição do ciclo celular em células HaCaT e SCC-9 / SCC-4 tratadas com 1, 5 e 10 $\mu\text{g} / \text{ml}$ de AG por 24, 48 e 72 h (25).

2. OBJETIVOS

2.1. Objetivo geral

Analisar o potencial antineoplásico da Nanopartícula Alumínio Cloro Ftalocianina e do Ácido Gálico.

2.2. Objetivos específicos

- Avaliar a atividade antineoplásica da nanopartícula Alumínio Cloro Ftalocianina e sua ação sobre a viabilidade das células de Carcinoma Epidermoide de Boca;
- Avaliar a eficácia do Ácido Gálico em modelos *in vivo* mediante avaliação clínica e histopatológica de camundongos *swiss* com tumores induzidos por administração 4nqo.

3. PRODUTOS

3.1. Produto 1:

Photodynamic therapy mediated by nanoparticles Aluminum Chloro Phthalocyanine in oral squamous carcinoma cells

Lilian Mendes Borburema Cangussu¹, Ludmilla Regina de Souza¹, Marcela Gonçalves de Souza, Luis Alexandre Muehlmann², Paulo Narcizo de Souza^{3,4}, Alfredo Maurício Batista de Paula¹, Lucyana Conceição Farias¹, Sérgio Henrique Sousa Santos⁵, and André Luiz Sena Guimarães^{1,6#}

¹Department of Dentistry, Universidade Estadual de Montes Claros (Unimontes), Montes Claros, Minas Gerais, Brazil

²Institute of Physics of the University of Brasilia; Faculty of Ceilandia, University of Brasilia, Federal District, Brazil

³Department of Genetics and Morphology, Institute of Biological Sciences, University of Brasilia, Federal District, Brazil

⁴Department of Physics, University of Brasilia, Federal District, Brazil

⁵Institute of Agricultural Sciences, Universidade Federal de Minas Gerais (UFMG), Montes Claros, Minas Gerais, Brazil

⁶Dilson Godinho Hospital, Montes Claros, Minas Gerais, Brazil

Corresponding author:

André Luiz Sena Guimarães. Laboratório de Pesquisa em Saúde, Hospital Universitário Clemente Faria, Universidade Estadual de Montes Claros, 562 Av. Cula Mangabeira Santo Expedito, Montes Claros, Minas Gerias, 39401-001 Brazil. E-mail: andreluizguimaraes@gmail.com

ABSTRACT

Objective: oral squamous cell carcinoma (OSCC) causes significant morbidity and mortality. The current study aims to investigate the antineoplastic potential of Photodynamic Therapy with photosensitizing (PDT) with Chloro-aluminum phthalocyanine (AICIPc) in OSCC.

Methods: A basic, applied cell study was performed. Four study groups were used Group 1: Control (did not receive any treatment), Group 2: PBS + 28.3Jcm² (received irradiation only), Group 3: AICIPc + 28.3Jcm² (received the photosensitizer and irradiation) and Group 4: AICIPc (received alone the photosensitizer). Initially, the concentration of AICIPc was tested. To test the effect of PDT with AICIPc migration and cell deaths were performed. Also, the immunoexpression of Ki-67 and TP53 was performed.

Results: All AICIPc concentrations (0.7, 0.035 and 0.017nM) significantly induced OSCC cell death. The lowest concentration was chosen for the tests. Migration and Cell death assays reveal that PDT with AICIPc significantly reduced migration and increased cell death when compared to controls. Also, PDT with AICIPc reduced Ki-67 and mutated TP53 immunoexpression.

Conclusion: PDT with AICIPc proved to be effective in altering the phenotype of OSCC reducing migration and inducing cell death. Additionally, PDT with AICIPc decreased Ki-67 and muted TP53 expression.

Keywords: Photodynamic Therapy. 4nqo. Antineoplastic action.

INTRODUCTION

Cancer burden rises to 18.1 million new cases and 9.6 million deaths (26). Oral squamous cell carcinoma (OSCC) reached 354,864 cases and the 177,384 deaths in the world during 2018 (27). Brazil is among the highest rates of OSCC in Latin America (28, 29). Most of OSCC cases commit a man. However, the gender difference in OSCC rates has been decreasing over time, presumably as tobacco and alcohol consumption equalize (30). The main risk factors for OSCC development are tobacco and alcohol (31), the use of which explains 82.9% of cases (32). Oral cancer causes significant morbidity and mortality (29). Moreover, OSCC is associated with significant effects on physiological function, facial esthetics, and survival (33).

The ultimate goal of OSCC treatment is to establish effective preventive and therapeutic measures against cancer, to preserve or to restore form and function, to minimize sequelae of treatment, and ultimately to prevent subsequent further cancers (34, 35). The currently available treatment modalities to OSCC include surgery, radiotherapy, chemotherapy, combined modality treatments and primary and secondary prevention strategies including lifestyle changes as well as chemoprevention (36).

Studies have demonstrated the use of alternative therapies in the control of OSCC (10, 34, 35, 37). Photodynamic therapy (PDT) is one of these alternatives to chemotherapy and radiotherapy for the treatment of malignant neoplasm (11, 12) because due to its local effects that make treatment less aggressive to the patient. PDT works by combining a photosensitizer drug (PS) with irradiation using visible light at an appropriate wavelength, which is individually harmless (38). The PS should have a high absorption peak between 600 and 800 nanometers (nm) (39). Thus, wavelengths longer than 800 nm does not provide enough energy to excite oxygen to its singlet state and to form a significant amount of reactive oxygen species (39).

Under irradiation, PSs transfer the absorbed light to the surrounding molecular oxygen, and cause cytotoxic reactive oxygen species (ROS) such as $^1\text{O}_2$ or free radicals, thus leading to cell apoptosis and destruction of the diseased tissue (40). The efficacy of a PDT depends on the formulation of drugs, which must accumulate selectively within tumor tissues in comparison to normal ones (41). PDT can promote

direct cytotoxic effects on tumor cells (39). Additionally, PDT might damage to the tumor vasculature, and induct a robust inflammatory reaction that can lead to the development of systemic immunity against the neoplasm (39).

Light emitting diode (LED) arrays are an effective and less costly means of light delivery (42). Developments in LED technology have provided higher power and narrower spectral characteristics making them a desirable alternative to lasers (43). The region between 600 and 1200 nm is generally referred to as a woven optical window (39). The photodynamic effects that occur after the absorption of visible light lead to the production of reactive oxygen species (ROS), which act as cytotoxic agents that can inactivate tumor cells (38). In the presence of oxygen, two competing reactions of the excited sensitizer can occur: a singlet or, more commonly, a triplet, can either react with the substrate or solvent (Type I) or with oxygen (Type II) (44).

Phthalocyanine derivatives comprise the second generation of photosensitizer molecules employed in PDT and have attracted much attention due to their outstanding photosensitizing performance (41). Chloro-aluminum phthalocyanine (AICIPc) is a chemically stable photosensitizer that displays photophysical properties that are highly suitable for PDT, such as high triplet quantum yields and long triplet lifetime (45). The system composed by AICIPc presents photodynamic activity in aqueous media and may be used for anticancer PDT (46). Studies have shown the efficient antitumor activity of AICIPc (46). To analyze the antineoplastic potential of the nanoparticle Aluminum Chlorine Phthalocyanine and its action on the viability of the cells of the in oral squamous carcinoma cells. This current study aims to investigate the antineoplastic potential of the AICIPc in OSCC.

MATERIAL AND METHODS

Study Design

A basic, applied cell study was performed to test the effect of PDT with AICIPc in OSCC cells.

AICIPc Concentration

Three AICIPc concentrations were tested 0.7, 0.035 and 0.017nM. The excitation was performed at 660 nm wavelength (fluency 28.35 Jcm² and power density 7.2 mW/cas) described before (47, 48).

LED source

All irradiations were performed under the following conditions: by 10 minutes, 05 cm away, wavelength 660 nm, 28.35 Jcm² and power density: 7.2 mW/cm², in an apparatus developed by Professor Paulo E. N. Souza, Prototype n.02. The light source used was composed of 20 LED light-emitting diodes (LED) model XL001WP01NRC660 (Shenzhen Sealand Optoelectronics Co., Ltd., China). The power circuit was assembled under the supervision of Prof. Dr. Paulo Souza of the Institute of Physics of the University of Brasília. The control of the power supply able to guarantee the stability of the lighting was carried out by an RCD-24-0.35 / W controller (Constant Current LED Driver, Recom Power, Inc., Germany). The emission spectrum of the LEDs was determined using a portable spectrophotometer (Ocean Optics Inc., USA). A digital potentiometer (Fieldmax II, Coherent, USA) was used to define the maximum power value of the LEDs, as well as the power values as a function of distance and the relative position of the LED bank to guarantee the reproducibility of the luminous intensity. Periodically the energy per unit of time was measured through the use of a potentiometer. The measurements recorded by the potentiometer and the detector area allowed determining that the light source has a power density of 0.04 W / cm². The LED apparatus used in this study was developed by Prof. Dr. Paulo Narcizo de Souza, at the University of Brasília, with low-cost

components and optimized for use in biological treatments (41) with the beginning of operation on 02/01/2015 - INCT / CNPQ University of Brasília, Institute of Physics.

Cells and experimental conditions

The Scc9 (Code. 0196) cells lines *Homo sapiens*; Tongue; Epithelial-Like were, and Cells A431, epidermal carcinoma, both cells were acquired at the Bank of Cells of Rio de Janeiro (BCRJ). Cells were cultured in DMEM/Ham's F-12 supplemented with 10% fetal bovine serum and 0.4 µg/ml hydrocortisone (Gibco, USA, Catalog number: 10437028), at 37°C with 5% CO₂ in a humidified air atmosphere. All experiments were performed in triplicate. The cells lines (2×10^6) were seeded in a 6-well plate supplemented with 10% FBS and incubated 24 h.

Cell proliferation

The cells of all groups were counted using a Neubauer chamber, 24 h after treatment. All groups were performed in triplicate. Cell proliferation assay was performed as described before (47) with necessary adaptations. A density of 2×10^5 OSCC cells was plated in 60 mm dish and incubated at 37 °C for approximately 24h to establish adherent monolayers. Then, cells were treated with the AICIPc concentration was previously defined through in the MTT assay.

Study Groups

The cells were distributed in four groups.

Group 1: Control (did not receive any type of treatment);

Group 2: PBS + 28.3Jcm² (received irradiation only);

Group 3: AICIPc + 28.3Jcm² (received the photosensitizer and irradiation) and

Group 4: AICIPc (received only the photosensitizer).

MTT

The cell viability was checked through assay. Cells were seeded in 96-well plate at the density of 3×10^3 cells/well and left in complete media containing 10% FBS for 24h. The cells were divided into two groups (AICIPc and AICIPc + LED), and each group was treated with AICIPc for 15 min. All groups were washed with PBS. One of the groups received irradiation. Control did not. After washing, the control group was incubated with PBS while the AICIPc group received radiation, only then the cells were placed in a dark incubator for 24h.

Migration assay

Wound healing method was used to test by cell migration (49). OSCC cells were plated in 60 mm dish at a density of 2.5×10^5 cells and maintained in culture for 24h. Before treatment, to determine the cell migration, a lesion was created on the monolayer cells using a sterile 10 μ l pipette tip. All groups were photographed. The cells of the Control Group and AICIPc Group have not been irradiated. While the 28.35 Jcm² Group and AICIPc + 28.35Jcm² Group was irradiated for 10 min and placed back in the incubator. The changes in the distance between the lesion edges area were monitored at 24h with the new photography. The migration rate (%) as the distance that cells migrate regarding the original distance of the lesion.

Each wound area was measured by automatic analysis software Image J. The migration ability was expressed as a percentage. Images of the wounded cell monolayers were taken using an Olympus IX81 inverted microscope (Olympus, Center Valley, PA, USA) coupled to camera SC30 (Olympus, Center Valley, PA, USA) before and at 24h after wounding.

Acridine Orange / Ethidium Bromide assay

Cell viability was analyzed in the Acridine orange / Ethidium bromide (AO/EB) assay staining used to visualize dead and viable cells (50). Pit by pit, the cells (3×10^4 cells/mL) were removed from the greenhouse and had their means replaced

by a volume 20 μ L of a solution containing 1 part of 100 μ g/mL Acridine orange in PBS; (AO, Sigma, St. Louis, USA) and 1 part of 100 μ g/mL Ethidium bromide in PBS (EB, Sigma, St. Louis, USA). Cells were observed in a fluorescence microscope FSX100 (Olympus, Center Valley, PA, USA).

Immunocytochemical assay

Immunocytochemical was performed as described before (49) with necessary adaptations. A density of 1×10^5 OSCC cells was plated 60mm culture plates and submitted to the assays. Then the cells were fixed with 70% ethanol for 30 min. Endogenous peroxidase activity was blocked with 0.3% hydrogen peroxide. The following primary mouse monoclonal antibodies were used: anti-Ki-67 (1:100, SolA15, Sigma, St. Louis, MO, USA) and TP53 (1:20, PAb 122, Invitrogen). All monoclonal antibodies were incubated for 18h at 4°C. Endogenous peroxidase was blocked by incubation with 0.03% H₂O₂ in ethanol for 30 min. The primary antibodies against Ki-67 and TP53 were detected using the Universal HRP Immunostaining Kit (KP-500, Diagnostic BioSystems, Pleasanton, CA, USA). Signals were developed with 3'3-diaminobenzidine-tetrahydrochloride for 5 min and counterstained with Mayer's hematoxylin for 30 sec. Negative controls were performed by replacing the primary antibody with PBS. Slides were photographed on Brightfield microscope FSX100 (Olympus, Center Valley, PA, USA) at 20 \times . The manual counts were performed in merge image by ImageJ software (51). Immunocytochemistry analyses of investigated antigens were carried out by determining the percentage of positively stained viable cells in all fields counted.

The numbers of positively stained and nonstained cells were recorded separately. A negative result was defined as the absence of stained and feasible nuclei. Staining was considered positive when brown nuclear labeling was observed.

RESULTS

Chloro-aluminum phthalocyanine concentration

The effectiveness of AICIPc as a photosensitizer with and without irradiating light emitting diode (LED) was further investigated using MTT (3- (4,5-dimethylthiazol-2-yl) -2,5-diphenyltetrazolium bromide). We have observed that PDT exhibits a significant anticancer effect on 24-hour MTT treatment. We confirm that the ALCIPc acts on the viability of OSCC cells at all concentrations tested (Figure 1).

After this assay, we decide to choose only one concentration for the other experiments. We chose the lowest dose of 0.017nM because it is a minimum dose of AICIPc (among those tested) capable of producing a therapeutic effect.

PDT reduce cell migration and induced cell death

Thus, the migration assay results indicated that the combined treatment of the AICIPc + 28.3 Jcm² considerably reduced cell migration compared with single treatments and PBS combined with LED (Figure 2).

Acridine orange penetrates living and dead cells, emitting green fluorescence as a result of the intercalation in the double-stranded DNA and red-orange fluorescence after binding with single-stranded RNA, while ethidium bromide emits red fluorescence after intercalation in the DNA of cells with altered cell membrane (at necrosis or a late stage of apoptosis) (52). Acridine Orange/Ethidium Bromide Cell death assay reveals that PDT significantly increased the number of death process cells when compared to controls (Figure 3).

Cellular development inhibition was observed when compared with controls, as can be seen in Fig. 3 OSCC cells with orange dots in cell nuclei treated with PDT. The maximum increase ($p < 0.05$) in the quantity of not viable cells was observed in treated cells. Low cytotoxicity was observed when cells were treated only with AICIPc and only LED with/or PBS.

PDT reduced Ki-67 and muted TP53 immunoeexpression

Some molecules affect the process of transformation of OSCC (28, 53). Among these molecule TP53 and Ki-67, are known to significantly influence in malignization of oral potentially malignant disorders (54). OSCC cells presented a robust expression of the proliferation marker Ki-67 positive cells (Figure 3). PDT with AICIPc at 28.3 J/cm² group drastically reduced Ki-67 (Figure 3). Muted TP53 is associated with OSCC worse prognoses (29, 55). Our results demonstrated that the expression of the mutated TP53 protein was dramatically reduced by PDT treatment (Figure 4). Additionally, a clear hypotrophy is observed OSCC cells treated with PDT.

DISCUSSION

OSCC is the most common cancer of the head and neck (26). Moreover, even with surgical reconstruction procedures, after OSCC treatment the occurrence of some degree of facial disfigurement, leading to severe aesthetic and functional defects (56). The current study presents a possible perspective to treat OSCC with AICIPc PDT.

Migration is a cyclic process in which the cell changes shape produces morphological asymmetry and translocates the cell body implying death in cancer patients related to metastatic progression (57). Increased migration at the individual cell level affects local neoplasm invasion and metastasis (67). In the current study, AICIPc mediated PDT significantly reduced cell migration. The reduction of migration is essential for local growth control (58).

Previous studies have performed AO/EB staining and reported that early apoptotic cells had fragmented DNA which exhibited strong green colored nuclei (34, 35). Dual AO/EB fluorescent staining can detect fundamental morphological changes in apoptotic cells. Besides, it allows for the distinction between healthy cells, early and late apoptotic cells, and necrotic cells (34, 35). The viable and non-apoptotic cells are stained green while the apoptotic cells are stained orange or red (59). In the current study, the OSCC cells treated only with light doses 28.3 J/cm² were non-toxic to OSCC cells. These results are by previous demonstrations that both the light source and drug carrier are innocuous when used separately (46, 52). On the other hand, our data demonstrated that PDT with AICIPc induced cell death. Moreover, all concentrations used in the current study (0.7, 0.035 and 0.017nM) caused cell death.

Our data corroborate with evidence that demonstrated that PDT could lead to all three forms of cell death (apoptosis, necrosis, and autophagy) by different signaling pathways (60) acting induced death by arresting cell cycle progression (38).

The protein Ki-67 is only produced in actively dividing cells; it is located in the nucleus on the structure that contains most of the cell's DNA (2). The levels of this protein and location vary through the cell cycle (61) and the expression of Ki-67 is shown in all stages, except G₀, whereas resting cells entering from G₀ lack Ki-67 in the early part of G₁(62). The level of Ki-67 expression has been used as a prognostic determination index in human cancers (54) for having variable value for detecting malignant potential (54, 63). Ki-67 scoring is essential for diagnosis for tumor grade, based on proportional of tumor-positive cells has usually used as an indication for evaluation (64). Uncontrolled proliferation is a common feature of malignant cells, therapeutic agents that target Ki-67 may be useful tools in cancer treatment (65). The over-proliferation is a crucial feature of tumor progression, and the high rate is related to lousy prognosis (66, 67). Our results showed that the PDT with AICIPc drastically reduced expression of this marker about the control groups. Also, as well as Ki-67, the TP53 expression index increased as the severity of lesion increased showing that there is a relation between these two markers (28-30, 54, 55). TP53 is a tumor suppressor gene located on chromosome 17p and mutations in this gene is one of the most common events in human carcinogenesis (68). Mutations or deletions in the TP53 gene are present in nearly 50% of human cancers (69). Mutations (70, 71) or biochemical changes (72) that cause loss of function plays an essential role in the development of OSCC. Lost of TP53 function impossibilities the proliferation control resulting in the occurrence of tumors (72). Besides this TP53 mutation probably renders more aggressive phenotype (73). These mutations occur in the expression of full-length TP53 protein, but the loss of wild type tumor suppressive function (74). Our results demonstrated that Photodynamic Therapy mediates by AICIPc caused the death of cells expressing the TP53 mutant protein thus reducing its expression. In conclusion, PDT with AICIPc proved to be effective in altering the phenotype of OSCC reducing migration and inducing cell death. Additionally, PDT with AICIPc decreased Ki-67 and muted TP53 expression.

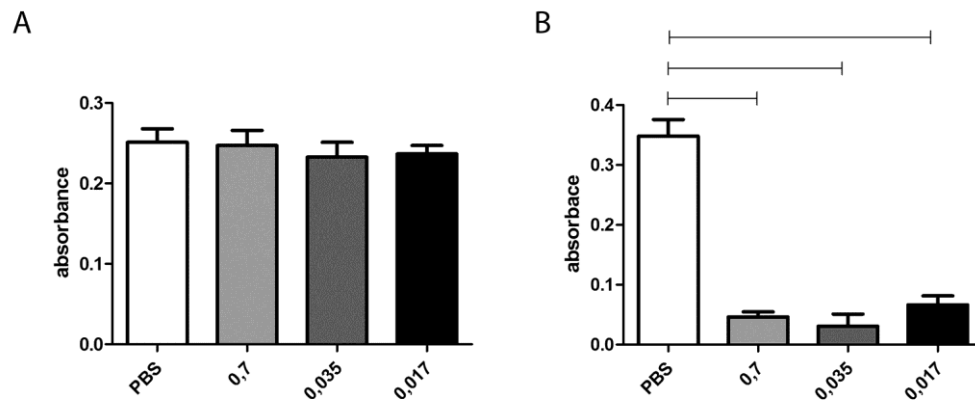


Figure 1.

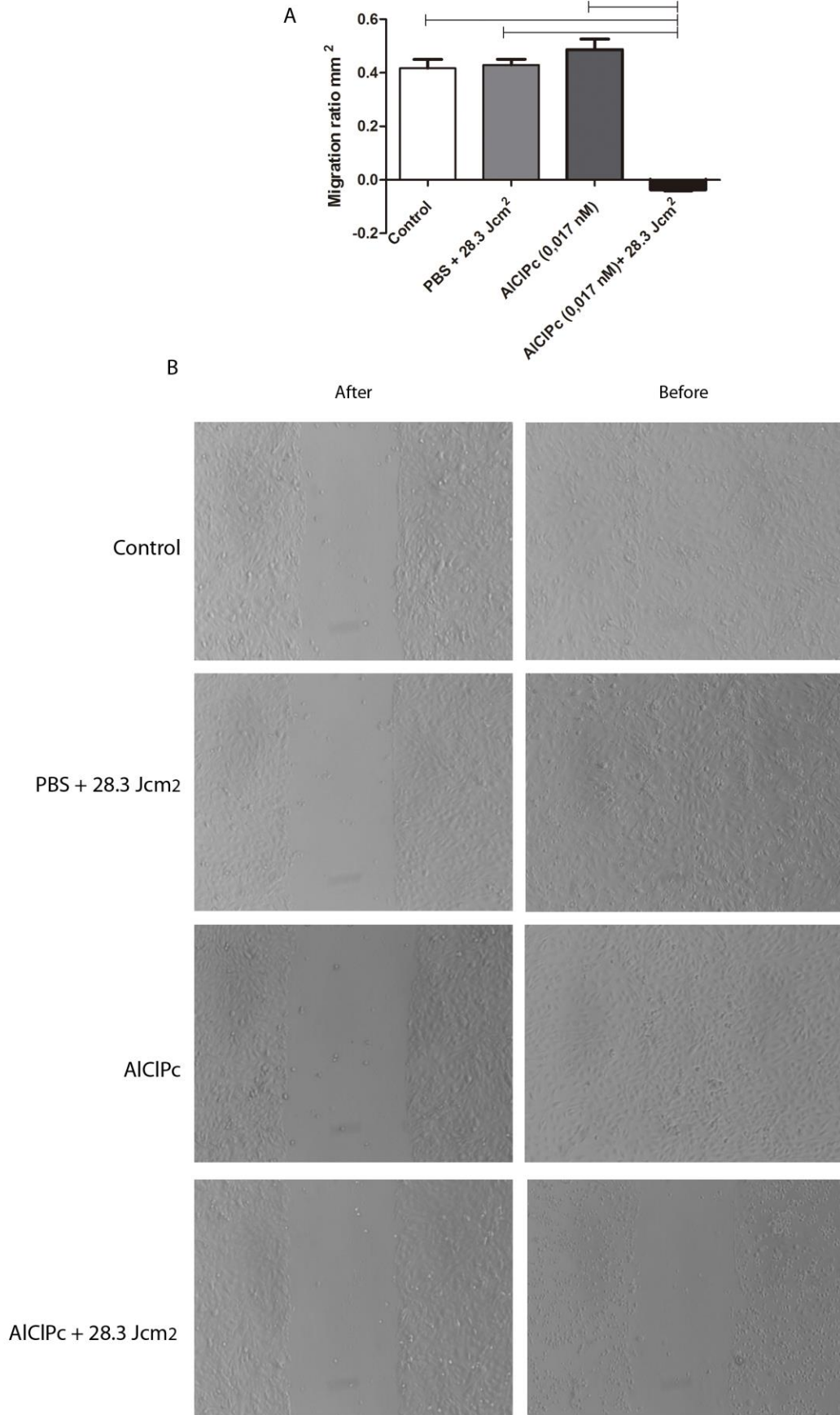


Figure 2.

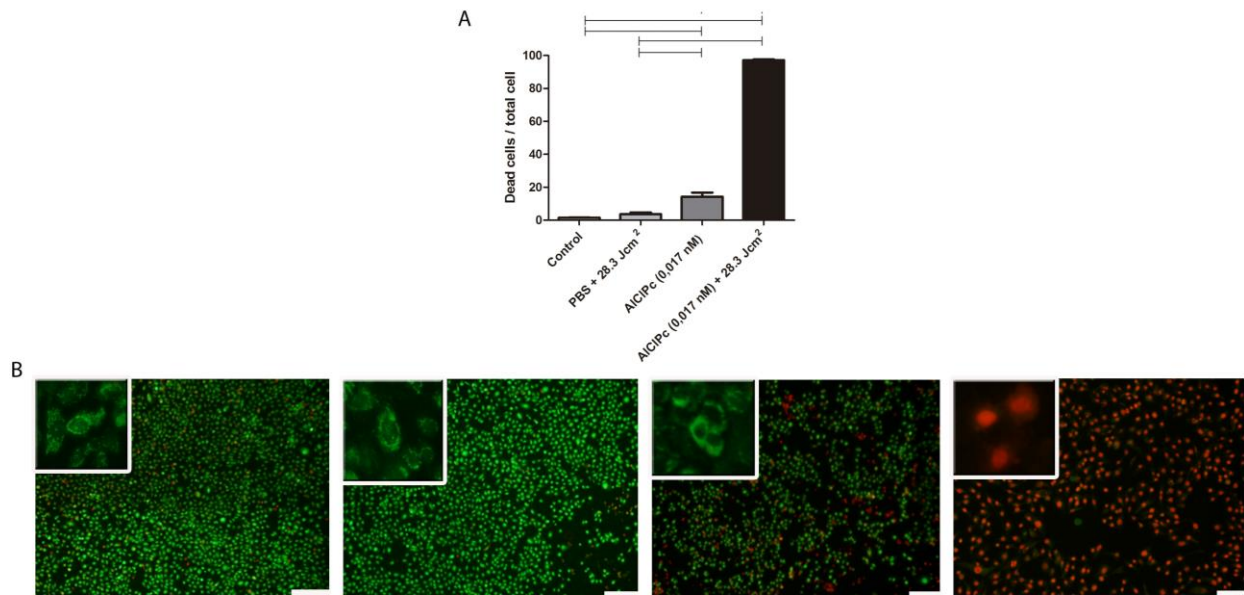


Figure 3.

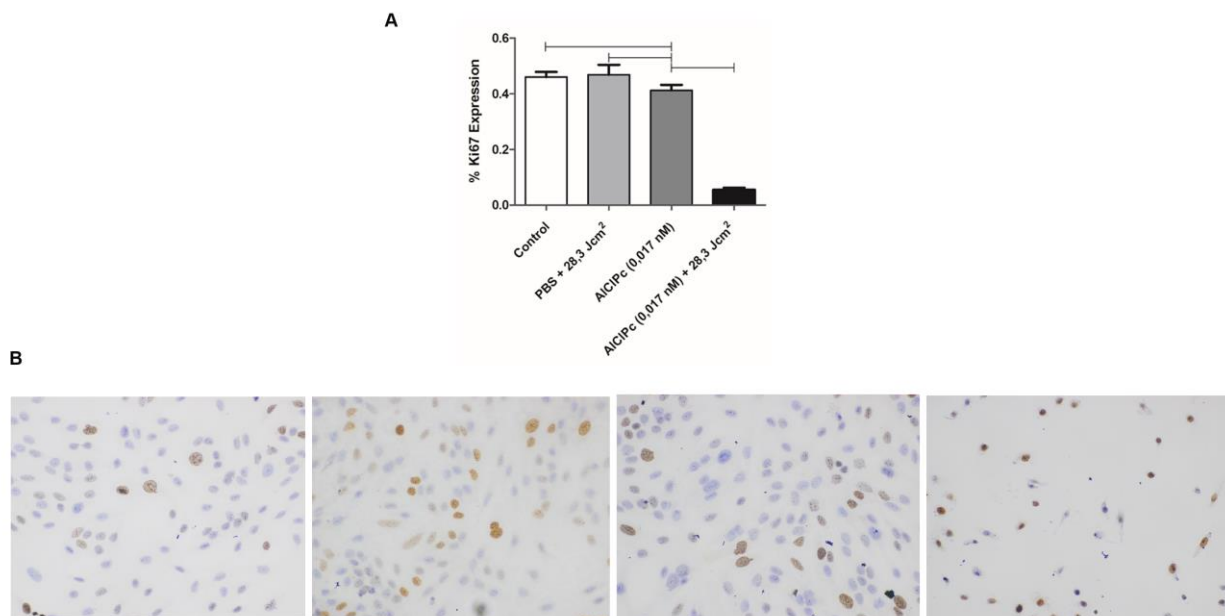


Figure 4.

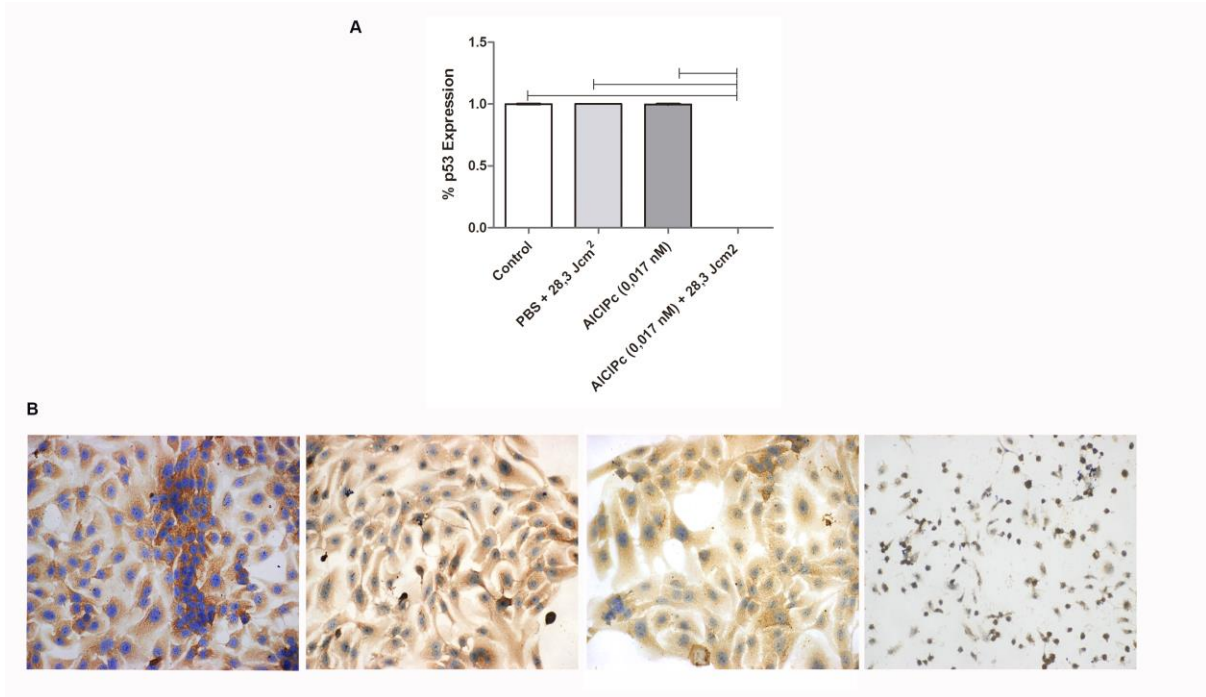


Figure 5.

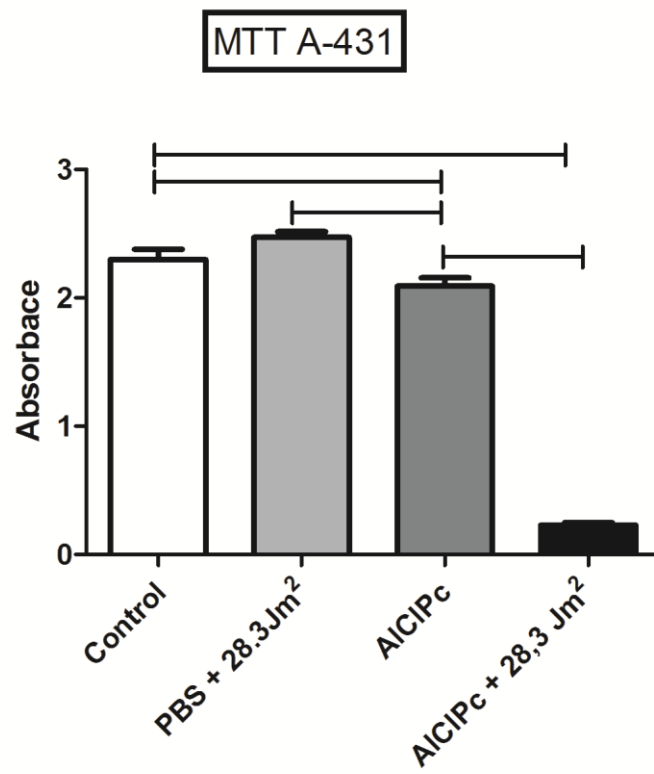


Figure 6.

Figure Legends

Figure 1. PDT suppresses the viability of SCC9 cells according to the MTT assay result. (A) Photodynamic therapy performed with PBS + LED 28.3 Jcm² (control). (B) Photodynamic therapy performed with AICIPc + LED (28.3 Jcm²). The horizontal bars represent a statistical difference between the bars that express the data.

Figure 2. Effect of PDT on cell migration. Quantification of the impact of PDT on the SCC9 cells (A). PDT drastically reduced cell migration ratio (B). The horizontal bars represent a statistical difference between the bars that express the data.

Figure 3. Effect of PDT on cell death. AO/EB representative figures (A) and quantification (B) show an increase in cell death as a consequence of PDT treatment. The horizontal bars represent a statistical difference between the bars that express the data.

Figure 4. Effect of PDT on cell death. (A) Ki-67 expression (B) Ki-67 expression show a decrease in expression this marker as a consequence of PDT treatment. The horizontal bars represent a statistical difference between the bars that express the data.

Figure 5. Effect of PDT on cell death. (A) TP53 expression (B) TP53 expression show a decrease in expression these markers as a consequence of PDT treatment. The horizontal bars represent a statistical difference between the bars that express the data.

Figure 6. PDT suppresses the viability of A-431 cells according to the MTT assay result. The horizontal bars represent a statistical difference between the bars that express the data.

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1. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: a cancer journal for clinicians*. 2018;68(6):394-424.
2. International Agency for Research on Cancer (IARC) [Internet]. 2018 [cited 18/11/2018].
3. Fraga CA, de Oliveira MV, de Oliveira ES, Barros LO, Santos FB, Gomez RS, et al. A high HIF-1alpha expression genotype is associated with poor prognosis of upper aerodigestive tract carcinoma patients. *Oral Oncol*. 2012;48(2):130-5.
4. De Paula AM, Souza LR, Farias LC, Correa GT, Fraga CA, Eleuterio NB, et al. Analysis of 724 cases of primary head and neck squamous cell carcinoma (HNSCC) with a focus on young patients and p53 immunolocalization. *Oral Oncol*. 2009;45(9):777-82.
5. Farias LC, Fraga CA, De Oliveira MV, Silva TF, Marques-Silva L, Moreira PR, et al. Effect of age on the association between p16CDKN2A methylation and DNMT3B polymorphism in head and neck carcinoma and patient survival. *International journal of oncology*. 2010;37(1):167-76.
6. Righini CA, Karkas A, Morel N, Soriano E, Reyt E. [Risk factors for cancers of the oral cavity, pharynx (cavity excluded) and larynx]. *Presse medicale (Paris, France : 1983)*. 2008;37(9):1229-40.
7. Radoi L, Menvielle G, Cyr D, Lapôte-Ledoux B, Stücker I, Luce D, et al. Population attributable risks of oral cavity cancer to behavioral and medical risk factors in France: results of a large population-based case-control study, the ICARE study. *BMC Cancer*. 2015;15:827.
8. Min S-K, Choi SW, Ha J, Park JY, Won Y-J, Jung K-W. Conditional relative survival of oral cavity cancer: Based on Korean Central Cancer Registry. *Oral Oncology*. 2017;72:73-9.
9. Guimaraes TA, Farias LC, Santos ES, de Carvalho Fraga CA, Orsini LA, de Freitas Teles L, et al. Metformin increases PDH and suppresses HIF-1alpha under hypoxic conditions and induces cell death in oral squamous cell carcinoma. *Oncotarget*. 2016;7(34):55057-68.
10. Guimaraes TA, Farias LC, Fraga CA, Feltenberger JD, Melo GA, Coletta RD, et al. Evaluation of the antineoplastic activity of gallic acid in oral squamous cell carcinoma under hypoxic conditions. *Anti-cancer drugs*. 2016;27(5):407-16.
11. Shah JP, Gil Z. Current concepts in management of oral cancer – Surgery. *Oral Oncology*. 2009;45(4):394-401.
12. Lee HM, Patel V, Shyur LF, Lee WL. Copper supplementation amplifies the anti-tumor effect of curcumin in oral cancer cells. *Phytomedicine : international journal of phytotherapy and phytopharmacology*. 2016;23(12):1535-44.
13. de Souza MG, de Jesus SF, Santos EM, Gomes ESB, de Paulo Santiago Filho A, Santos EMS, et al. Radiation Therapy Reduced Blood Levels of LDH, HIF-1alpha, and miR-210 in OSCC. *Pathology oncology research : POR*. 2018.
14. Biel MA. Photodynamic therapy treatment of early oral and laryngeal cancers. *Photochemistry and photobiology*. 2007;83(5):1063-8.
15. Chen HM, Chen CT, Yang H, Lee MI, Kuo MY, Kuo YS, et al. Successful treatment of an extensive verrucous carcinoma with topical 5-aminolevulinic acid-mediated photodynamic therapy. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2005;34(4):253-6.
16. Castilho-Fernandes A, Lopes TG, Primo FL, Pinto MR, Tedesco AC. Photodynamic process induced by chloro-aluminum phthalocyanine nanoemulsion in glioblastoma. *Photodiagnosis and Photodynamic Therapy*. 2017;19:221-8.

17. Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, et al. Photodynamic therapy of cancer: an update. *CA: a cancer journal for clinicians*. 2011;61(4):250-81.
18. Huang Z, Xu H, Meyers AD, Musani AI, Wang L, Tagg R, et al. Photodynamic therapy for treatment of solid tumors--potential and technical challenges. *Technology in cancer research & treatment*. 2008;7(4):309-20.
19. Py-Daniel KR, Namban JS, de Andrade LR, de Souza PEN, Paterno LG, Azevedo RB, et al. Highly efficient photodynamic therapy colloidal system based on chloroaluminum phthalocyanine/pluronic micelles. *European Journal of Pharmaceutics and Biopharmaceutics*. 2016;103:23-31.
20. Schmidt MH, Meyer GA, Reichert KW, Cheng J, Krouwer HG, Ozker K, et al. Evaluation of photodynamic therapy near functional brain tissue in patients with recurrent brain tumors. *Journal of neuro-oncology*. 2004;67(1-2):201-7.
21. Quirk BJ, Brandal G, Donlon S, Vera JC, Mang TS, Foy AB, et al. Photodynamic therapy (PDT) for malignant brain tumors – Where do we stand? *Photodiagnosis and Photodynamic Therapy*. 2015;12(3):530-44.
22. Foote CS. Definition of type I and type II photosensitized oxidation. *Photochemistry and photobiology*. 1991;54(5):659.
23. Nunes SM, Sguilla FS, Tedesco AC. Photophysical studies of zinc phthalocyanine and chloroaluminum phthalocyanine incorporated into liposomes in the presence of additives. *Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas*. 2004;37(2):273-84.
24. Muehlmann LA, Ma BC, Longo JP, Almeida Santos Mde F, Azevedo RB. Aluminum-phthalocyanine chloride associated to poly(methyl vinyl ether-co-maleic anhydride) nanoparticles as a new third-generation photosensitizer for anticancer photodynamic therapy. *International journal of nanomedicine*. 2014;9:1199-213.
25. Muehlmann LA, Rodrigues MC, Longo JP, Garcia MP, Py-Daniel KR, Veloso AB, et al. Aluminium-phthalocyanine chloride nanoemulsions for anticancer photodynamic therapy: Development and in vitro activity against monolayers and spheroids of human mammary adenocarcinoma MCF-7 cells. *Journal of nanobiotechnology*. 2015;13:36.
26. Morgado LF, Travolo ARF, Muehlmann LA, Narcizo PS, Nunes RB, Pereira PAG, et al. Photodynamic Therapy treatment of onychomycosis with Aluminium-Phthalocyanine Chloride nanoemulsions: A proof of concept clinical trial. *Journal of photochemistry and photobiology B, Biology*. 2017;173:266-70.
27. Chan SW, Lim CJ, Guo K, Ng CP, Lee I, Hunziker W, et al. A role for TAZ in migration, invasion, and tumorigenesis of breast cancer cells. *Cancer research*. 2008;68(8):2592-8.
28. Kasibhatla S, Amarante-Mendes GP, Finucane D, Brunner T, Bossy-Wetzel E, Green DR. Acridine Orange/Ethidium Bromide (AO/EB) Staining to Detect Apoptosis. *Cold Spring Harbor Protocols*. 2006;2006(3):pdb.prot4493.
29. Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. *Nature methods*. 2012;9(7):671-5.
30. Castano AP, Demidova TN, Hamblin MR. Mechanisms in photodynamic therapy: part one—photosensitizers, photochemistry and cellular localization. *Photodiagnosis and photodynamic therapy*. 2004;1(4):279-93.
31. Fonseca-Silva T, Farias LC, Cardoso CM, Souza LR, Carvalho Fraga CA, Oliveira MV, et al. Analysis of p16(CDKN2A) methylation and HPV-16 infection in oral mucosal dysplasia. *Pathobiology : journal of immunopathology, molecular and cellular biology*. 2012;79(2):94-100.

32. Pereira CS, Oliveira MV, Fraga CA, Barros LO, Domingos PL, Roy A, et al. Impact of the epithelial dysplasia grading and Ki67 proliferation index in the adjacent non-malignant mucosa on recurrence and survival in head and neck squamous cell carcinoma. *Pathology, research and practice*. 2012;208(11):651-6.
33. de Oliveira MV, Pereira Gomes EP, Pereira CS, de Souza LR, Barros LO, Mendes DC, et al. Prognostic value of microvessel density and p53 expression on the locoregional metastasis and survival of the patients with head and neck squamous cell carcinoma. *Applied immunohistochemistry & molecular morphology : AIMM*. 2013;21(5):444-51.
34. Barclay CW, Foster EC, Taylor CL. Restorative aspects of oral cancer reconstruction. *British dental journal*. 2018;225(9):848-54.
35. Justus CR, Leffler N, Ruiz-Echevarria M, Yang LV. In vitro cell migration and invasion assays. *Journal of visualized experiments : JoVE*. 2014(88):51046.
36. Domingos PLB, Souza MG, Guimaraes TA, Santos ES, Farias LC, de Carvalho Fraga CA, et al. Hypoxia reduces the E-cadherin expression and increases OSCC cell migration regardless of the E-cadherin methylation profile. *Pathology, research and practice*. 2017;213(5):496-501.
37. Jabir MS, Taha AA, Sahib UI, Taqi ZJ, Al-Shammari AM, Salman AS. Novel of nano delivery system for Linalool loaded on gold nanoparticles conjugated with CALNN peptide for application in drug uptake and induction of cell death on breast cancer cell line. *Materials science & engineering C, Materials for biological applications*. 2019;94:949-64.
38. Debele TA, Peng S, Tsai H-C. Drug Carrier for Photodynamic Cancer Therapy. *International journal of molecular sciences*. 2015;16(9):22094-136.
39. Sobecki M, Mrouj K, Camasses A, Parisi N, Nicolas E, Llères D, et al. The cell proliferation antigen Ki-67 organises heterochromatin. *eLife*. 2016;5:e13722-e.
40. Sobecki M, Mrouj K, Camasses A, Parisi N, Nicolas E, Llères D, et al. The cell proliferation antigen Ki-67 organises heterochromatin. *eLife*. 2016;5:e13722.
41. Scholzen T, Gerdes J. The Ki-67 protein: from the known and the unknown. *Journal of cellular physiology*. 2000;182(3):311-22.
42. Jing Y, Yang Y, Hao F, Song Y, Zhang X, Zhang Y, et al. Higher Ki67 expression in fibroblast like cells at invasive front indicates better clinical outcomes in oral squamous cell carcinoma patients. *Bioscience reports*. 2018;38(6).
43. Hasegawa T, Yamamoto S, Yokoyama R, Umeda T, Matsuno Y, Hirohashi S. Prognostic significance of grading and staging systems using MIB-1 score in adult patients with soft tissue sarcoma of the extremities and trunk. *Cancer*. 2002;95(4):843-51.
44. Kausch I, Lingnau A, Endl E, Sellmann K, Deinert I, Ratliff TL, et al. Antisense treatment against Ki-67 mRNA inhibits proliferation and tumor growth in vitro and in vivo. *International journal of cancer*. 2003;105(5):710-6.
45. Li C-F, Chen L-T, Lan J, Chou F-F, Lin C-Y, Chen Y-Y, et al. AMACR amplification and overexpression in primary imatinib-naïve gastrointestinal stromal tumors: a driver of cell proliferation indicating adverse prognosis. *Oncotarget*. 2014;5(22):11588-603.
46. Mohamed AA, Abbas MY, Alharbi H, Babiker AY. Assessment of Expression of Ki-67 in Benign and Malignant Prostatic Lesions among Sudanese Patients. *Open Access Macedonian Journal of Medical Sciences*. 2018;6(10):1809-12.
47. Zhu J, Sammons MA, Donahue G, Dou Z, Vedadi M, Getlik M, et al. Prevalent p53 mutants co-opt chromatin pathways to drive cancer growth. *Nature*. 2015;525(7568):206-11.
48. Ozaki T, Nakagawara A. Role of p53 in Cell Death and Human Cancers. *Cancers*. 2011;3(1):994-1013.
49. Pinheiro UB, Fraga CA, Mendes DC, Farias LC, Cardoso CM, Silveira CM, et al. Fuzzy clustering demonstrates that codon 72 SNP rs1042522 of TP53 gene associated with

HNSCC but not with prognoses. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2015;36(12):9259-65.

50. Souza LR, Fonseca-Silva T, Pereira CS, Santos EP, Lima LC, Carvalho HA, et al. Immunohistochemical analysis of p53, APE1, hMSH2 and ERCC1 proteins in actinic cheilitis and lip squamous cell carcinoma. *Histopathology*. 2011;58(3):352-60.

51. Pereira T, Brito JAR, Guimaraes ALS, Gomes CC, de Lacerda JCT, de Castro WH, et al. MicroRNA profiling reveals dysregulated microRNAs and their target gene regulatory networks in cemento-ossifying fibroma. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2018;47(1):78-85.

52. Hientz K, Mohr A, Bhakta-Guha D, Efferth T. The role of p53 in cancer drug resistance and targeted chemotherapy. *Oncotarget*. 2017;8(5):8921-46.

53. Chan JY-H, Chen Y-C, Liu S-T, Chou W-Y, Ho C-L, Huang S-M. Characterization of a new mouse p53 variant: loss-of-function and gain-of-function. *Journal of biomedical science*. 2014;21(1):40-.

3.2 Produto 2:

Chemopreventive effects of Gallic Acid on oral carcinogenicity induced by 4NQO in the tongue of mice

Lílian Mendes Borburema Cangussu¹, Ludmilla Regina de Souza¹, Eliane Macedo Sobrinho Santos², Marcela Gonçalves de Souza¹, Luís Paulo de Oliveira¹, Lincoln Valério Andrade Rodrigues¹, Carlos Ícaro de Jesus Silva¹, Guilherme Machado Xavier⁴, Alfredo Maurício Batista de Paula¹, Lucyana Conceição Farias¹, Sérgio Henrique Sousa Santos³, and André Luiz Sena Guimarães^{1,5#}

¹Department of Dentistry, Universidade Estadual de Montes Claros (Unimontes), Montes Claros, Minas Gerais, Brazil

² Instituto Federal do Norte de Minas Gerais - Campus Araçuaí, Minas Gerais, Brazil

³Institute of Agricultural Sciences, Universidade Federal de Minas Gerais (UFMG), Montes Claros, Minas Gerais, Brazil

⁴Department of Orthodontics, King's College London Dental Institute, London, UK

⁵Dilson Godinho Hospital, Montes Claros, Minas Gerais, Brazil

Corresponding author:

André Luiz Sena Guimarães. Laboratório de Pesquisa em Saúde, Hospital Universitário Clemente Faria, Universidade Estadual de Montes Claros, 562 Av. Cula Mangabeira Santo Expedito, Montes Claros, Minas Gerais, 39401-001 Brazil. E-mail: andreluizguimaraes@gmail.com

ABSTRACT

Objective: As oral cancer development is a complex process, animal models represent an essential alternative to evaluate OSCC potential treatments. As such, the current study aims to investigate the GA preventive effect and potential toxicity.

Methods: The animals were kept in an environment with a controlled temperature of $21 \pm 2^{\circ}\text{C}$ and with a cycle of 12 hours of light /12 hours of dark (lights on from 12 hours, and they were fed with rations and filtered water. Animals were followed from birth to 234 days. The animals were randomly divided into four groups. A total of 39 mice *Mus musculus* males were distributed into four groups as follows: - Group 1 (Control): negative control group (N=7); - Group 2 (GA): treated with Gallic Acid (n=7); - Group 3 (GA+4NQO): received 4NQO and Gallic Acid treatment (n=7); - Group 4 (4NQO): received only 4NQO (n=7)

Results: Absent/mild epithelial dysplasia; moderate/severe epithelial dysplasia and in situ/invasive carcinoma distribution were different among groups. GA treatment statistically reduced the histopathological grade of 4NQO induced lesions. Also, GA did not promote epithelial changes in the absence of 4 NQO. Statistical differences in survival were also observed among the groups. In the presence of 4NQO, GA treatment increased the survival. Also, GA did not promote animal deaths in the absence of 4NQO.

Conclusion: AC reduced the severity of OSCC lesions, increased survival and reduced 4NQO liver toxicity in mice and decreased the mice weight loss in consequence of 4NQO.

Keywords: Antioxidant effect. Antineoplastic action. Polyphenols. Oral squamous cell carcinoma

INTRODUCTION

Oral Squamous Cell Carcinoma (OSCC) is worldwide common malignant neoplasm (75). OSCC main etiologically factor is tobacco consumption especially in association with alcohol (29, 76). Recently the potential role of oncogenic HPV has been extensively discussed in the literature (53, 77). Biochemical and molecular changes also can change OSCC development and prognosis (28, 30, 53, 70, 78-80). A critical issue in OSCC prevention is the absence of a deterministic biomarker to identify OSCC development (81, 82). As oral cancer development is a complex process, animal models represent an essential alternative to evaluate OSCC potential treatments (83, 84).

Experimental carcinogenesis induced by synthetic chemical 4-nitroquinoline 1-oxide (4NQO) is one of the most frequently used methods in animals for the study of OSCC (85). 4NQO is a synthetic water-soluble chemical carcinogen that forms all similar to the genetic alterations provoked by tobacco carcinogens (86). Mice exposed to 4NQO exhibit 1 to 3 carcinomas together with multiple dysplastic lesions at the end of the carcinogen exposure, thus limiting the ability to examine the benefit of preventive agents unless they also display anticancer properties (87). The tongue carcinogenesis model parallels the development of tongue cancer in humans as the dysplastic lesions are produced by long-term ingestion of small amounts of carcinogen (88). Moreover, 4NQO for 16 weeks cause hepatic and renal toxicity (89). Currently, the treatment of OSCC is surgery, radiation, and chemotherapy alone or combined (90). Treatment strategies for OSCC vary based on the stage at the time of diagnosis (90). Patients with the localized disease typically receive surgery and radiotherapy, leading to a high probability of long-term survival but with considerable morbidity (91). Simultaneously chemotherapy and radio therapy have given significant beneficial responses at inoperable stages of the disease (92). Other potential OSCC treatments emerged, like an EGFR Inhibitors: Tyrphostin AG-1478 that improves (cisplatin sensitivity of the OSCC (93) and Nimotuzumab in combination with PDT (photodynamic therapy) that increase the antitumor effect of PDT in an oral tumor model (94). But despite all improvements in OSCC treatment, the survival of OSCC patients population remains around 50% in five years (29, 75, 77).

Currently, OSCC treatment and prevention represents a challenge to the health systems (95, 96). Significant reduction in OSCC advantage stages is achieved by a population-based screening program (97). However, a considerable decrease in OSCC incidence is not obtained only with only early detection programs (96). Moreover, completely quit smoking is difficult to achieve especially in patients with low social support, depression, greater nicotine dependence and poor social-emotional function (98). A decrease in smoking behavior was observed with multidisciplinary smoking cessation advisor team but, after 5-year follow-up appointments, 74% of patients ultimately continued to smoke (99).

Considering the difficulty in achieving the quit smoking, new therapies to minimize smoke effects and prevent OSCC are very important (81). Nimesulide decreased the incidence of tongue lesions on Fisher 344, rats with induced squamous cell carcinomas (100); but as a selective COX-2 inhibitor might represent critical adverse effects in clinical practice (101). Metformin is also a potential agent to treat OSCC (34, 102) who presents clinical risks (103).

Gallic acid (GA), 3,4,5-trihydroxybenzoic acid, ($C_6H_2(OH)_3COOH$) is one of the polyphenolic substances in plants (104). In nature, GA are present in nearly every part of the plant, such as bark, wood, leaf, fruit, root, and seed (104). They are present in different concentrations in common foodstuffs such as blueberry, blackberry, strawberry, plums, grapes, mango, cashew nut, hazelnut, walnut, tea, wine (105). Another important GA source is the Pequi (*Caryocar brasiliense* Camb.)(106). GA inhibits triggers apoptosis through the regulation of Fas/FasL, p53, and Bcl-2 family and activation of caspase cascade (107). Moreover, studies demonstrated that GA selectively induces OSCC cell death by apoptosis (108), even under hypoxia (35). GA also decrease of invasiveness (109) inhibits of NF-B (110) and have anti-angiogenesis activities (111).

The cytotoxicity shown by GA is not a frequent feature in phenolic compounds but is a reasonably specific characteristic of GA that contains three adjacent phenolics, hydroxyl groups were responsible for the cytotoxicity, and the carboxyl group that is implicated in distinguishing between normal cells and cancer cells (112). Preclinical studies to evaluate the GA toxicity and preventive effect in OSCC are scarce, despite the potential of GA to treat OSCC (35). As such, the current study aims to investigate the preventive effect of GA and potential toxicity.

MATERIALS AND METHODS

Ethical approval for this study was obtained from the relevant Institutional Animal Care and Use Committee (protocol 098/16).

Animals and experimental conditions

Where used in this study mice males swiss (*Mus musculus*) received food and water *ad libitum*. The animals were kept in an environment with a controlled temperature of $21 \pm 2^{\circ}\text{C}$ and with a cycle of 12 hours of light /12 hours of dark (lights on from 12 hours, and they were fed with rations and filtered water. Animals were followed from birth to 234 days.

Study Groups

Animals were randomly divided into four groups. A total of 39 mice *Mus musculus* males were distributed into four groups as follows:

- Group 1 (Control): negative control group (N=7)
- Group 2 (GA): treated with Gallic Acid (n=7)
- Group 3 (GA+4NQO): received 4NQO and Gallic Acid treatment (n=7)
- Group 4 (4NQO): received only 4NQO (n=7)

Cancer Induction

Induction of oral carcinogenesis was performed using 4-nitroquinoline-1-oxide (4NQO; N8141-1G, Sigma-Aldrich, St. Louis, USA) in drinking water to a final concentration of $100\mu\text{g/ml}$ as described before (113). After 39 day-olds weighing at the weight of 20 to 22 g mice received the treatment with 4NQO for 112 days. Animals were followed from birth to until the death or 234TH day.

Treatment

The establishment of Gallic Acid was according to our and other previous data (108, 113, 114). The daily administration of Gallic Acid (100 mg/Kg) by oral gavage started at the day 67th until the 234th-day lingual tissue of the animals.

End of the experiment

The endpoint of the experiment was the animal death, or at the 234th day, the remaining animals were sacrificed via decapitation through a guillotine.

Animal Macroscopic and weight evaluation

A general macroscopic evaluation (115) and body weight (116) evaluation were performed, and the groups were compared.

Histological preparation and analyses

Shortly after their sacrifices, all organs were placed in properly labeled containers and was fixed in 10% formalin solution for 48 hours. All histological techniques were performed following the protocol previously described (54, 117). Briefly, tissues were included in paraffin. The slides were stained with hematoxylin-eosin samples were covered with glass coverslips for observation and histological quantification through microscopy (Olympus Fsx100, Center Valley, Palo Alto, CA). Twenty fields of each tissue (oral mucosa, kidney, and liver) of each group were randomly photographed using a microscope (FSX100, Olympus, Center Valley, PA, USA) by a researcher who did not participate of the quantification or analyses. All quantifications were made using ImageJ software (118), which was used previously (108, 113). Histological specimens were blindly examined by a trained pathologist using the criteria recommended for histopathological diagnosis. The morphological semi-quantitatively analyses were performed in the liver and kidney to evaluate toxicity as described before (89). The OSCC lesions were graded according to previous studies (113, 119).

Statistical analysis

The PASW Statistics18-SPSS software (IBM, Armonk, NY) was used for the statistical analysis. Kolmogorov–Smirnov and Shapiro-Wilk tests were used in samples that had nonparametric distribution, before, were submitted to an independent T-test, and samples that did not follow this distribution were subjected to a Mann-Whitney nonparametric test. The LSD Test was used in relative animal weight investigations. Categorical variables were analyzed by chi-square or Fisher's exact. The Kaplan-Meier method was used in estimating survival relative to time, and survival differences were analyzed with the Log-rank test as done in SPSS. Confidence above 95% ($P < 0.05$) was considered to be significant statistical analysis showing. GraphPad Prism (GraphPad Software, San Diego, CA, USA) was used for in the construction of the graphs and $p < 0.05$ was considered statistically significant.

RESULTS

The Macroscopic features of mice were used to compare the effect of GA. 4NQO (Group 3 and 4) induced weight loss in comparison to control groups (Group 1 and 2). The GA treatment (Group 2) promote weight reduction, in contrast, to control (Group 1). On the other hand, in mice treated with 4NQO (Groups 3 and 4), GA induces an increase in body weight (Group 4) (Figure 1).

Absent/mild epithelial dysplasia; moderate/severe epithelial dysplasia and in situ/invasive carcinoma distribution were different among groups (Figure 2A). GA treatment statistically reduced the histopathological grade of 4NQO induced lesions (Group 4, Figure 2A). Also, GA did not promote epithelial changes in the absence of 4 NQO (Groups 1 and 2, Figure 2A). Statistical differences in survival were also observed among the groups (Figure 2B). In the presence of 4NQO (Groups 3 and 4), GA treatment increased the survival (Group 4, Figure 2B). Also, GA did not promote animal deaths in the absence of 4NQO (Group 2, Figure 2B).

The histopathological characterization of the liver tissue of the animals in the four groups of the present study are disposed of in figure 3. 4NQO treatment (group 3)

present liver distinct cellular and histopathological changes such as steatosis in the microvascular pattern (Figure 3A). GA promote attenuation of 4NQO damage in kidney tissue (Figure 3B).

DISCUSSION

As the treatment of early staged OSCC patients is associated with higher quality of life survival (91), OSCC needs to be imminent diagnosed and treated (95-97). But OSCC patients have still been diagnosed and treated in advanced stages (29, 75, 77). Moreover, there is no preventive treatment for OSCC development in patients exposed OSCC risk factors (81). Also, no even drug could delay OSCC development in patients presented OSCC risk factors (81). The necessity to at least promote a delay in OSCC development might help to improve the local control and survival (120). Weight loss is a critical phenomenon that commits OSCC patients (121). In the absence of 4NQO GA treatment promote weight loss. It is observed that the activation of the AMPK/Sirt1/PGC1 α pathway are GA target and consequently made GA a potential therapeutic intervention for insulin resistance in metabolic diseases (122) and could explain the result of the current study in group 2. Moreover, GA also reduced the weight of animals by lowering plasma glucose and triglyceride levels in addition to reducing the weight of adipose peri-renal tissues of rats (123). However, in the current study, when mice were submitted to 4NQO GA presents opposite effect in body weight. GA promoted the reduction of weight loss in group 4. It was demonstrated that GA changes OSCC reduced HIF-1 α expression (35). HIF-1 α is also directed related to the glycolytic pathway in the OSCC context (34). The dual effect of GA might be explained by the capacity of GA restore normal metabolism, specifically related to the glycolytic pathway, shifting to aerobic glycolysis (124). Moreover, the molecules related to metabolic syndrome such as angiotensinogen converting enzyme (ACE) might be associated with OSCC (81).

The current study demonstrated that GA attenuates the severity of the mice oral lesions. Recent studies have shown that GA can induce apoptosis as a result of the anti-inflammatory activity (125) by intrinsic mitochondrial pathway (126, 127). GA has also presented effects of DNA methyltransferases activities suggesting that GA might be useful dietary intervention strategy for tobacco-associated cancers (128). Additionally, GA treatment resulted in significant changes in cell membrane lipids and

fatty acids inducing malignant cells to apoptosis pathways (129). These findings corroborate the current study that demonstrated that systemic administration of GA has chemopreventive activity during oral carcinogenesis induced by 4NQO. Benefits of GA are perceived for other cancers such as gastric and lung cancer (130, 131). Curcumin (132) and Pecan nut shell extract, which are also rich in phenolic compounds, showed antitumor activity against MCF-7 human breast cancer cells, decreasing the viability of the cells, increasing the cell death by apoptosis and arresting cell cycle (133).

The systemic 4NQO toxicity is well established (89). GA supplementation also represents systemic benefits (114, 124).

GA provide adequate protection against oxidative stress damage in the liver in rats reducing lipid peroxidation and free radical levels and increasing enzymatic and non-enzymatic antioxidant defense in these tissues (134). At the same dose of the current study (100 mg/kg), GA is effective against chronic liver damage, induced by the administration of fluoxetine (114). GA treatment also prevented the increase in reactive species in the liver and kidney of diabetic rats (134). In corroboration with findings of the current study indicate GA was not only well tolerated but also reduced 4NQO toxicity.

The limitation of the current study is related to its design. Preclinical studies should be confirmed in humans. On the other hand for the first time, GA was tested effect and toxicity in an OSCC established model.

In conclusion: AC reduced the severity of OSCC lesions, increased survival and reduced 4NQO liver toxicity in mice. Additionally AC reduced the mice weight loss in consequence of 4NQO.

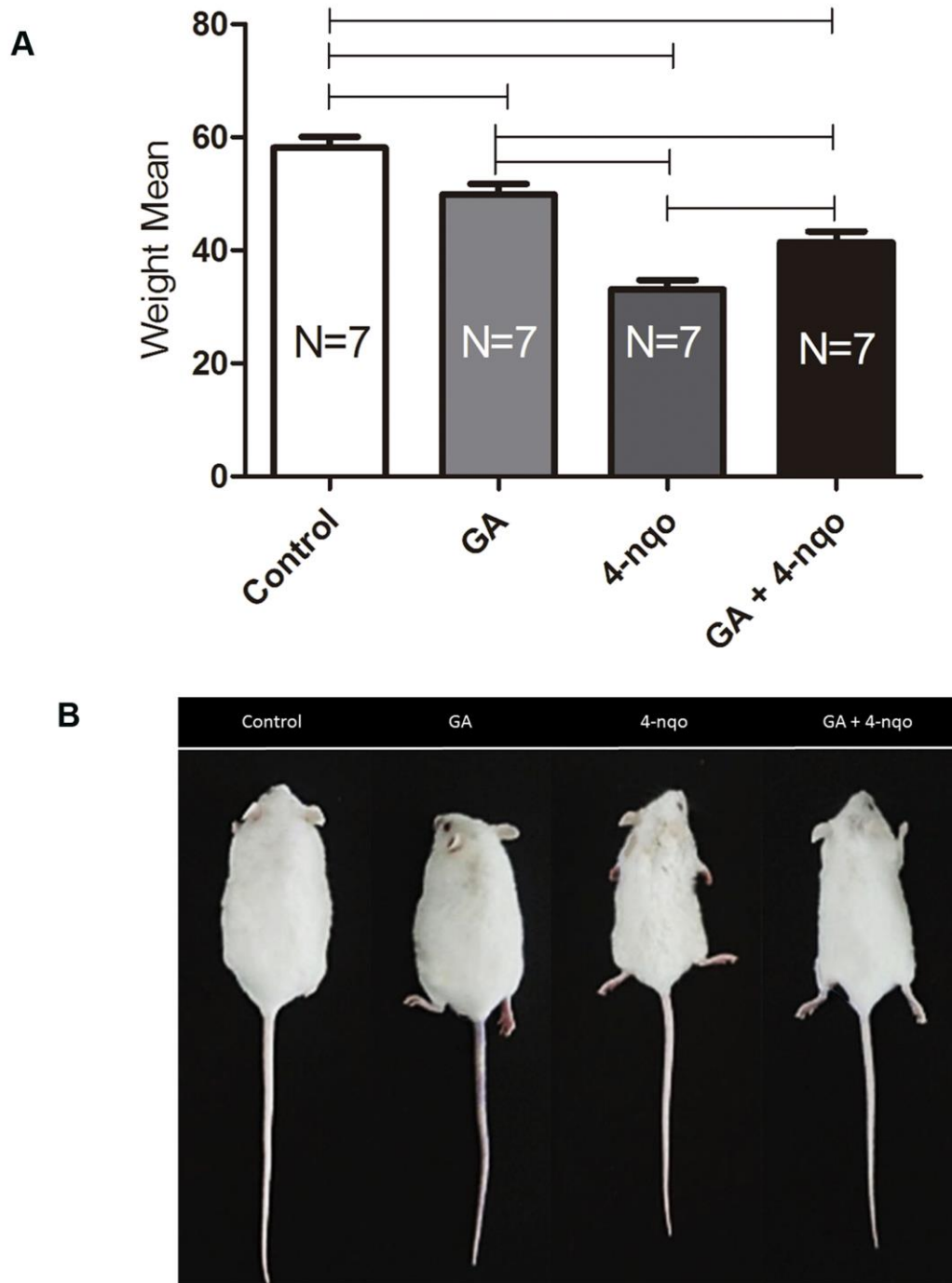
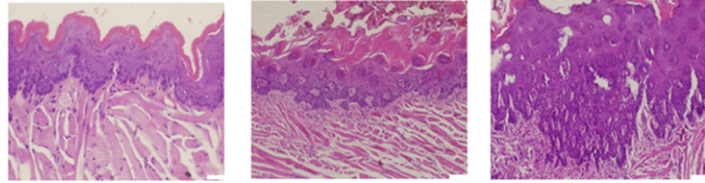


Figure 1.

A



	Dysplasia Absent/Mild	Dysplasia Moderate/Severe	Carcinoma	<i>p</i> value
Control	7	0	0	
GA	7	0	0	
4NQO	1	3	3	
GA + 4NQO	3	3	1	0.006

B

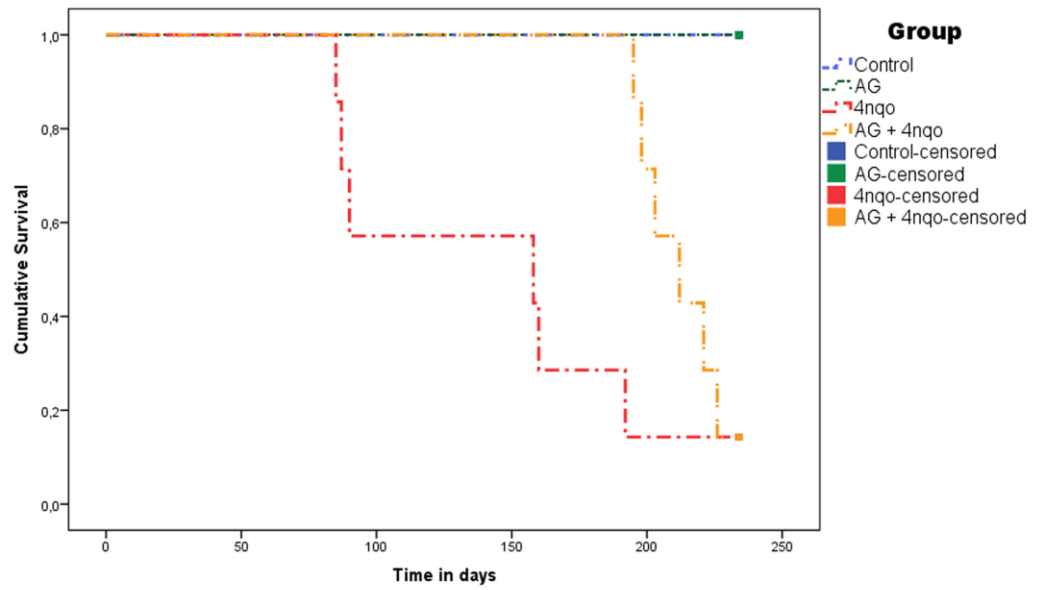


Figure 2.

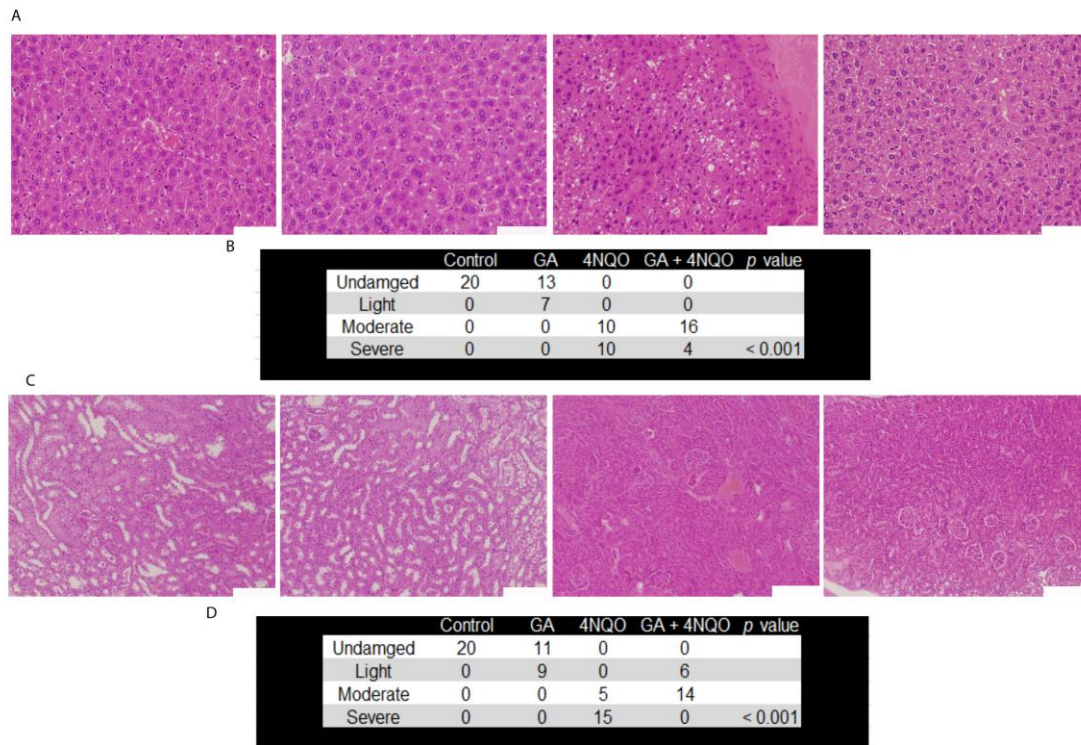


Figure 3.

Figure Legends

Figure 1. Effect of Gallic Acid in animals' weight

Weight average between groups of animals. Mice treated with 4NQO (Group 3 and 4) had lower body weight than the control groups (Group 1 and 2) ($p < 0.05$). The treatment with GA provided gain in body weight compared to the group receiving only 4NQO. However, treated animals (Group 3) did not reach the same weight level as the control group (Group 1). The horizontal bars represent a statistical difference ($p < 0.05$) between the bars that express the data.

Figure 2: Effect of Gallic Acid in the incidence of Oral cancer and mice survival

In A, Gallic acid reduced the severity of 4NQO treatment in the histological grading ($p = 0.006$). Scale bar represent 64 μm . The rate of mortality was significantly lower in mice receiving 4NQO and GA than mice that received only 4NQO ($p < 0.05$) in the Kaplan-Meier survival (B).

Figure 3 Effect of GA under the expected 4NQO in the animals' livers (A). Gallic acid alone did not cause hepatic tissue damage. Moreover, Gallic acid also reduced 4NQO liver toxicity ($p < 0.05$). Similarly, in kidney Gallic acid also protected against 4NQO toxicity (B, $p < 0.05$). Scale bar represent 10 μm .

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1. Shield KD, Ferlay J, Jemal A, Sankaranarayanan R, Chaturvedi AK, Bray F, et al. The global incidence of lip, oral cavity, and pharyngeal cancers by subsite in 2012. *CA: a cancer journal for clinicians*. 2017;67(1):51-64.
2. Souza RL, Fonseca-Fonseca T, Oliveira-Santos CC, Correa GT, Santos FB, Cardoso CM, et al. Lip squamous cell carcinoma in a Brazilian population: epidemiological study and clinicopathological associations. *Med Oral Patol Oral Cir Bucal*. 2011;16(6):e757-62.
3. De Paula AM, Souza LR, Farias LC, Correa GT, Fraga CA, Eleuterio NB, et al. Analysis of 724 cases of primary head and neck squamous cell carcinoma (HNSCC) with a focus on young patients and p53 immunolocalization. *Oral oncology*. 2009;45(9):777-82.
4. Marques-Silva L, Farias LC, Fraga CA, de Oliveira MV, Cardos CM, Fonseca-Silva T, et al. HPV-16/18 detection does not affect the prognosis of head and neck squamous cell carcinoma in younger and older patients. *Oncology letters*. 2012;3(4):945-9.
5. Fonseca-Silva T, Farias LC, Cardoso CM, Souza LR, Carvalho Fraga CA, Oliveira MV, et al. Analysis of p16(CDKN2A) methylation and HPV-16 infection in oral mucosal dysplasia. *Pathobiology : journal of immunopathology, molecular and cellular biology*. 2012;79(2):94-100.
6. Farias LC, Fraga CA, De Oliveira MV, Silva TF, Marques-Silva L, Moreira PR, et al. Effect of age on the association between p16CDKN2A methylation and DNMT3B polymorphism in head and neck carcinoma and patient survival. *International journal of oncology*. 2010;37(1):167-76.
7. Correa GT, Bandeira GA, Cavalcanti BG, de Carvalho Fraga CA, dos Santos EP, Silva TF, et al. Association of -308 TNF-alpha promoter polymorphism with clinical aggressiveness in patients with head and neck squamous cell carcinoma. *Oral oncology*. 2011;47(9):888-94.
8. Fraga CA, de Oliveira MV, de Oliveira ES, Barros LO, Santos FB, Gomez RS, et al. A high HIF-1alpha expression genotype is associated with poor prognosis of upper aerodigestive tract carcinoma patients. *Oral oncology*. 2012;48(2):130-5.
9. de Carvalho Fraga CA, Farias LC, de Oliveira MV, Domingos PL, Pereira CS, Silva TF, et al. Increased VEGFR2 and MMP9 protein levels are associated with epithelial dysplasia grading. *Pathology, research and practice*. 2014;210(12):959-64.
10. Pinheiro UB, de Carvalho Fraga CA, Mendes DC, Marques-Silva L, Farias LC, de Souza MG, et al. p16 (CDKN2A) SNP rs11515 was not associated with head and neck carcinoma. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2014;35(6):6113-8.
11. Pinheiro UB, Fraga CA, Mendes DC, Farias LC, Cardoso CM, Silveira CM, et al. Fuzzy clustering demonstrates that codon 72 SNP rs1042522 of TP53 gene associated with HNSCC but not with prognoses. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2015;36(12):9259-65.
12. de Carvalho Fraga CA, Farias LC, Jones KM, Batista de Paula AM, Guimaraes ALS. Angiotensin-Converting Enzymes (ACE and ACE2) as Potential Targets for Malignant Epithelial Neoplasia: Review and Bioinformatics Analyses Focused in Oral Squamous Cell Carcinoma. *Protein and peptide letters*. 2017;24(9):784-92.
13. van der Waal I. Oral potentially malignant disorders: is malignant transformation predictable and preventable? *Med Oral Patol Oral Cir Bucal*. 2014;19(4):e386-90.
14. Vairaktaris E, Spyridonidou S, Papakosta V, Vylliotis A, Lazaris A, Perrea D, et al. The hamster model of sequential oral oncogenesis. *Oral oncology*. 2008;44(4):315-24.
15. Liu X, He R, Chen W. [A rat model of tongue mucosa squamous cell carcinoma induced by oral administration of 4NQO in drinking water]. *Zhonghua kou qiang yi xue za zhi = Zhonghua kouqiang yixue zazhi = Chinese journal of stomatology*. 1999;34(6):354-6.

16. Dayan D, Hirshberg A, Kaplan I, Rotem N, Bodner L. Experimental tongue cancer in desalivated rats. *Oral oncology*. 1997;33(2):105-9.
17. Vitale-Cross L, Czerninski R, Amornphimoltham P, Patel V, Molinolo AA, Gutkind JS. Chemical carcinogenesis models for evaluating molecular-targeted prevention and treatment of oral cancer. *Cancer prevention research (Philadelphia, Pa)*. 2009;2(5):419-22.
18. Vitale-Cross L, Molinolo AA, Martin D, Younis RH, Maruyama T, Patel V, et al. Metformin prevents the development of oral squamous cell carcinomas from carcinogen-induced premalignant lesions. *Cancer prevention research (Philadelphia, Pa)*. 2012;5(4):562-73.
19. Nagler R, Dayan D. The dual role of saliva in oral carcinogenesis. *Oncology*. 2006;71(1-2):10-7.
20. Barcessat AR, Huang I, Rabelo GD, Rosin FC, Ferreira LG, de Cerqueira Luz JG, et al. Systemic toxic effects during early phases of topical 4-NQO-induced oral carcinogenesis in rats. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2014;43(10):770-7.
21. Mendenhall WM, Dagan R, Bryant CM, Fernandes RP. Radiation Oncology for Head and Neck Cancer: Current Standards and Future Changes. *Oral and maxillofacial surgery clinics of North America*. 2019;31(1):31-8.
22. Algazi AP, Grandis JR. Head and neck cancer in 2016: A watershed year for improvements in treatment? *Nature reviews Clinical oncology*. 2017;14(2):76-8.
23. Lasrado S, Moras K, Pinto GJ, Bhat M, Hegde S, Sathian B, et al. Role of concomitant chemoradiation in locally advanced head and neck cancers. *Asian Pacific journal of cancer prevention : APJCP*. 2014;15(10):4147-52.
24. Hiraishi Y, Wada T, Nakatani K, Tojyo I, Matsumoto T, Kiga N, et al. EGFR inhibitor enhances cisplatin sensitivity of oral squamous cell carcinoma cell lines. *Pathology oncology research : POR*. 2008;14(1):39-43.
25. Bhuvanewari R, Ng QF, Thong PS, Soo KC. Nimotuzumab increases the anti-tumor effect of photodynamic therapy in an oral tumor model. *Oncotarget*. 2015;6(15):13487-505.
26. Sankaranarayanan R, Mathew B, Jacob BJ, Thomas G, Somanathan T, Pisani P, et al. Early findings from a community-based, cluster-randomized, controlled oral cancer screening trial in Kerala, India. The Trivandrum Oral Cancer Screening Study Group. *Cancer*. 2000;88(3):664-73.
27. Frenandez Garrote L, Sankaranarayanan R, Lence Anta JJ, Rodriguez Salva A, Maxwell Parkin D. An evaluation of the oral cancer control program in Cuba. *Epidemiology*. 1995;6(4):428-31.
28. Chuang SL, Su WW, Chen SL, Yen AM, Wang CP, Fann JC, et al. Population-based screening program for reducing oral cancer mortality in 2,334,299 Taiwanese cigarette smokers and/or betel quid chewers. *Cancer*. 2017;123(9):1597-609.
29. Chang SL, Lo CH, Peng HL, Chen CR, Wu SC, Chen SC. Factors associated with continued smoking after treatment of oral cavity cancer: An age and survival time-matched study. *Journal of advanced nursing*. 2018;74(4):926-34.
30. Hamadah O, Hepburn S, Thomson PJ. Effects of active non-smoking programmes on smoking behaviour in oral precancer patients. *International journal of oral and maxillofacial surgery*. 2007;36(8):706-11.
31. Yamamoto K, Kitayama W, Denda A, Morisaki A, Kuniyasu H, Kirita T. Inhibitory effects of selective cyclooxygenase-2 inhibitors, nimesulide and etodolac, on the development of squamous cell dysplasias and carcinomas of the tongue in rats initiated with 4-nitroquinoline 1-oxide. *Cancer letters*. 2003;199(2):121-9.

32. Donati M, Conforti A, Lenti MC, Capuano A, Bortolami O, Motola D, et al. Risk of acute and serious liver injury associated to nimesulide and other NSAIDs: data from drug-induced liver injury case-control study in Italy. *British journal of clinical pharmacology*. 2016;82(1):238-48.
33. Guimaraes TA, Farias LC, Santos ES, de Carvalho Fraga CA, Orsini LA, de Freitas Teles L, et al. Metformin increases PDH and suppresses HIF-1alpha under hypoxic conditions and induces cell death in oral squamous cell carcinoma. *Oncotarget*. 2016;7(34):55057-68.
34. Cabrera E, Levenson J, Armentano R, Barra J, Pichel R, Simon AC. Aortic pulsatile pressure and diameter response to intravenous perfusions of angiotensin, norepinephrine, and epinephrine in conscious dogs. *Journal of cardiovascular pharmacology*. 1988;12(6):643-9.
35. Lalau JD. Lactic acidosis induced by metformin: incidence, management and prevention. *Drug safety*. 2010;33(9):727-40.
36. Subramanian AP, Jaganathan SK, Mandal M, Supriyanto E, Muhamad II. Gallic acid induced apoptotic events in HCT-15 colon cancer cells. *World journal of gastroenterology*. 2016;22(15):3952-61.
37. Daglia M, Di Lorenzo A, Nabavi SF, Talas ZS, Nabavi SM. Polyphenols: well beyond the antioxidant capacity: gallic acid and related compounds as neuroprotective agents: you are what you eat! *Current pharmaceutical biotechnology*. 2014;15(4):362-72.
38. de Oliveira TS, Thomaz DV, da Silva Neri HF, Cerqueira LB, Garcia LF, Gil HPV, et al. Neuroprotective Effect of Caryocar brasiliense Camb. Leaves Is Associated with Anticholinesterase and Antioxidant Properties. *Oxidative medicine and cellular longevity*. 2018;2018:9842908-.
39. Verma S, Singh A, Mishra A. Gallic acid: molecular rival of cancer. *Environmental toxicology and pharmacology*. 2013;35(3):473-85.
40. Santos EMS, da Rocha RG, Santos HO, Guimaraes TA, de Carvalho Fraga CA, da Silveira LH, et al. Gallic acid modulates phenotypic behavior and gene expression in oral squamous cell carcinoma cells by interfering with leptin pathway. *Pathology, research and practice*. 2018;214(1):30-7.
41. Guimaraes TA, Farias LC, Fraga CA, Feltenberger JD, Melo GA, Coletta RD, et al. Evaluation of the antineoplastic activity of gallic acid in oral squamous cell carcinoma under hypoxic conditions. *Anti-cancer drugs*. 2016;27(5):407-16.
42. Lu Y, Jiang F, Jiang H, Wu K, Zheng X, Cai Y, et al. Gallic acid suppresses cell viability, proliferation, invasion and angiogenesis in human glioma cells. *European journal of pharmacology*. 2010;641(2-3):102-7.
43. Ho HH, Chang CS, Ho WC, Liao SY, Lin WL, Wang CJ. Gallic acid inhibits gastric cancer cells metastasis and invasive growth via increased expression of RhoB, downregulation of AKT/small GTPase signals and inhibition of NF-kappaB activity. *Toxicology and applied pharmacology*. 2013;266(1):76-85.
44. Liu Z, Schwimer J, Liu D, Lewis J, Greenway FL, York DA, et al. Gallic acid is partially responsible for the antiangiogenic activities of Rubus leaf extract. *Phytotherapy research : PTR*. 2006;20(9):806-13.
45. Inoue M, Suzuki R, Sakaguchi N, Li Z, Takeda T, Ogihara Y, et al. Selective induction of cell death in cancer cells by gallic acid. *Biological & pharmaceutical bulletin*. 1995;18(11):1526-30.
46. Sobrinho Santos EM, Guimaraes TA, Santos HO, Cangussu LMB, de Jesus SF, Fraga CAC, et al. Leptin acts on neoplastic behavior and expression levels of genes related to hypoxia, angiogenesis, and invasiveness in oral squamous cell carcinoma. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2017;39(5):1010428317699130.

47. Karimi-Khouzani O, Heidarian E, Amini SA. Anti-inflammatory and ameliorative effects of gallic acid on fluoxetine-induced oxidative stress and liver damage in rats. *Pharmacological Reports*. 2017;69(4):830-5.
48. Han ED, MacFarlane RC, Mulligan AN, Scafidi J, Davis AE, 3rd. Increased vascular permeability in C1 inhibitor-deficient mice mediated by the bradykinin type 2 receptor. *The Journal of clinical investigation*. 2002;109(8):1057-63.
49. Mendes-Junior LG, Freitas-Lima LC, Oliveira JR, Melo MB, Feltenberger JD, Brandi IV, et al. The usefulness of short-term high-fat/high salt diet as a model of metabolic syndrome in mice. *Lasers in medical science*. 2018;209:341-8.
50. Gomes EP, Aguiar JC, Fonseca-Silva T, Dias LC, Moura-Boas KP, Roy A, et al. Diazepam reverses the alveolar bone loss and hippocampal interleukin-1beta and interleukin-6 enhanced by conditioned fear stress in ligature-induced periodontal disease in rats. *Journal of periodontal research*. 2013;48(2):151-8.
51. Pereira CS, Oliveira MV, Fraga CA, Barros LO, Domingos PL, Roy A, et al. Impact of the epithelial dysplasia grading and Ki67 proliferation index in the adjacent non-malignant mucosa on recurrence and survival in head and neck squamous cell carcinoma. *Pathology, research and practice*. 2012;208(11):651-6.
52. Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. *Nature Methods*. 2012;9:671.
53. Warnakulasuriya S, Reibel J, Bouquot J, Dabelsteen E. Oral epithelial dysplasia classification systems: predictive value, utility, weaknesses and scope for improvement. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2008;37(3):127-33.
54. Li C-C, Shen Z, Bavarian R, Yang F, Bhattacharya A. Oral Cancer: Genetics and the Role of Precision Medicine. *Dental Clinics of North America*. 2018;62(1):29-46.
55. Correa GT, Bandeira GA, Cavalcanti BG, Santos FB, Rodrigues Neto JF, Guimaraes AL, et al. Analysis of ECOG performance status in head and neck squamous cell carcinoma patients: association with sociodemographical and clinical factors, and overall survival. *Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer*. 2012;20(11):2679-85.
56. Doan KV, Ko CM, Kinyua AW, Yang DJ, Choi YH, Oh IY, et al. Gallic acid regulates body weight and glucose homeostasis through AMPK activation. *Endocrinology*. 2015;156(1):157-68.
57. Lucena SR, Salazar N, Gracia-Cazaña T, Zamarrón A, González S, Juarranz Á, et al. Combined Treatments with Photodynamic Therapy for Non-Melanoma Skin Cancer. *International journal of molecular sciences*. 2015;16(10):25912-33.
58. Chao J, Huo TI, Cheng HY, Tsai JC, Liao JW, Lee MS, et al. Gallic acid ameliorated impaired glucose and lipid homeostasis in high fat diet-induced NAFLD mice. *PloS one*. 2014;9(2):e96969.
59. Li D, Liu Z, Zhao W, Xi Y, Niu F. A straightforward method to determine the cytotoxic and cytopathic effects of the functional groups of gallic acid. *Process Biochemistry*. 2011;46(11):2210-4.
60. Sun G, Zhang S, Xie Y, Zhang Z, Zhao W. Gallic acid as a selective anticancer agent that induces apoptosis in SMMC-7721 human hepatocellular carcinoma cells. *Oncology Letters*. 2016;11(1):150-8.
61. Inoue M, Suzuki R, Koide T, Sakaguchi N, Ogihara Y, Yabu Y. Antioxidant, Gallic Acid, Induces Apoptosis in HL-60RG Cells. *Biochemical and Biophysical Research Communications*. 1994;204(2):898-904.

62. Weng Y-P, Hung P-F, Ku W-Y, Chang C-Y, Wu B-H, Wu M-H, et al. The inhibitory activity of gallic acid against DNA methylation: application of gallic acid on epigenetic therapy of human cancers. *Oncotarget*. 2018;9(1):361-74.
63. Rattanata N, Daduang S, Wongwattanakul M, Leelayuwat C, Limpiboon T, Lekphrom R, et al. Gold Nanoparticles Enhance the Anticancer Activity of Gallic Acid against Cholangiocarcinoma Cell Lines. *Asian Pacific journal of cancer prevention : APJCP*. 2015;16(16):7143-7.
64. Ho H-H, Chang C-S, Ho W-C, Liao S-Y, Lin W-L, Wang C-J. Gallic acid inhibits gastric cancer cells metastasis and invasive growth via increased expression of RhoB, downregulation of AKT/small GTPase signals and inhibition of NF- κ B activity. *Toxicology and applied pharmacology*. 2013;266(1):76-85.
65. You BR, Park WH. Gallic acid-induced lung cancer cell death is related to glutathione depletion as well as reactive oxygen species increase. *Toxicology in Vitro*. 2010;24(5):1356-62.
66. de Paiva Gonçalves V, Ortega AAC, Guimarães MR, Curylofo FA, Junior CR, Ribeiro DA, et al. Chemopreventive Activity of Systemically Administered Curcumin on Oral Cancer in the 4-Nitroquinoline 1-Oxide Model. *Journal of Cellular Biochemistry*. 2015;116(5):787-96.
67. Hilbig J, Policarpi PdB, Grinevicius VMAdS, Mota NSRS, Toaldo IM, Luiz MTB, et al. Aqueous extract from pecan nut [*Carya illinoensis* (Wangenh) C. Koch] shell show activity against breast cancer cell line MCF-7 and Ehrlich ascites tumor in Balb-C mice. *Journal of Ethnopharmacology*. 2018;211:256-66.
68. de Oliveira LS, Thome GR, Lopes TF, Reichert KP, de Oliveira JS, da Silva Pereira A, et al. Effects of gallic acid on delta - aminolevulinic dehydratase activity and in the biochemical, histological and oxidative stress parameters in the liver and kidney of diabetic rats. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2016;84:1291-9.

CONCLUSÕES

O objetivo principal deste trabalho foi avaliar o potencial antineoplásico da nanopartícula Alumínio Cloro Ftalocianina (AICIPc) e do Ácido Gálico (AG).

Os dois produtos propostos apresentaram resposta positiva nos experimentos até aqui realizados.

Ao serem avaliados os níveis de morte celular provocados pelo fotossensibilizante AICIPc, através da TFD, é possível inferir sobre a eficácia do procedimento proposto. Portanto, concluímos que a Terapia Fotodinâmica se mostrou eficiente no tratamento de OSCC ao ser realizada com a nanopartícula AICIPc.

Em nosso trabalho, o AG colaborou para que as lesões presentes nas línguas e fígado dos animais tivessem sua gravidade amenizada. Além disto, se mostrou eficiente na manutenção do peso e aparência dos animais, provocando considerável aumento de peso quando comparado com o grupo que recebeu apenas o carcinógeno. O Ácido Gálico apresentou capacidade de atenuar a gravidade das lesões apesar de não impedir a instalação das mesmas nas línguas dos animais tratados.

Estes resultados podem permitir que novos protocolos sejam desenvolvidos para estudos posteriores *in vivo*.

REFERÊNCIAS

1. Saúde Md. Estimativa Câncer de Boca Rio de Janeiro: Instituto Nacional do Câncer; 2018.
2. Sobocki M, Mrouj K, Camasses A, Parisi N, Nicolas E, Llères D, et al. The cell proliferation antigen Ki-67 organises heterochromatin. *eLife*. 2016;5:e13722-e. PubMed PMID: 26949251.
3. Saúde Md. Diretrizes da Política Nacional de Saúde Bucal. In: Bucal CNdS, editor. Brasília 2008.
4. Saúde Md. Caderno de Atenção Básica. In: Básica DdA, editor. Brasília 2008. p. 92.
5. LONGO JPF, LOZZI SP, AZEVEDO CB. Câncer bucal e a terapia fotodinâmica como modalidade terapêutica. *RGOR* revista Gaúcha de Odontologia (Online). 2011;59:51-7.
6. Oda D, Habte T, Yamachika E. Artemisinin: an alternative treatment for oral cancer. *Oral Surgery, Oral Medicine, Oral Pathology, Oral Radiology, and Endodontology*. 2004 2004/08/01;98(2):204.
7. Rahman MA, Amin ARM, Shin DM. Chemopreventive potential of natural compounds in head and neck cancer. *Nutrition and cancer*. 2010;62(7):973-87. PubMed PMID: 20924973.
8. Godsey J, Grundmann O. Review of Various Herbal Supplements as Complementary Treatments for Oral Cancer. *Journal of dietary supplements*. 2016;13(5):538-50. PubMed PMID: 26863913. Epub 2016/02/13. eng.
9. Ledwith K, Ogburn R, Cox J, Graham R, Fritzsche A, Gosnell D, et al. Taxol: efficacy against oral squamous cell carcinoma. *Mini reviews in medicinal chemistry*. 2013 Apr;13(4):509-21. PubMed PMID: 23373651. Epub 2013/02/05. eng.
10. Lee HM, Patel V, Shyur LF, Lee WL. Copper supplementation amplifies the anti-tumor effect of curcumin in oral cancer cells. *Phytomedicine : international journal of phytotherapy and phytopharmacology*. 2016 Nov 15;23(12):1535-44. PubMed PMID: 27765374. Epub 2016/10/22. eng.
11. Biel MA. Photodynamic therapy treatment of early oral and laryngeal cancers. *Photochemistry and photobiology*. 2007 Sep-Oct;83(5):1063-8. PubMed PMID: 17880501. Epub 2007/09/21. eng.
12. Chen HM, Chen CT, Yang H, Lee MI, Kuo MY, Kuo YS, et al. Successful treatment of an extensive verrucous carcinoma with topical 5-aminolevulinic acid-mediated photodynamic therapy. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2005 Apr;34(4):253-6. PubMed PMID: 15752262. Epub 2005/03/09. eng.
13. Vered M, Daniel N, Hirshberg A, Dayan D. Histomorphologic and morphometric changes in minor salivary glands of the rat tongue during 4-nitroquinoline-1-oxide-induced carcinogenesis. *Oral oncology*. 2003 2003/07/01;39(5):491-6.
14. Dayar D, Hirshberg A, Kaplan I, Rotem N, Bodner L. Experimental tongue cancer in desalivated rats. *Oral oncology*. 1997 1997/03/01;33(2):105-9.
15. Castilho-Fernandes A, Lopes TG, Primo FL, Pinto MR, Tedesco AC. Photodynamic process induced by chloro-aluminum phthalocyanine nanoemulsion in glioblastoma. *Photodiagnosis Photodyn Ther*. 2017 Sep;19:221-8. PubMed PMID: 28599959. Epub 2017/06/11. eng.
16. Moore JV. Photodynamic Therapy: Basic Principles and Clinical Applications. *British Journal of Cancer*. 1993;68(6):1257-. PubMed PMID: PMC1968635.
17. Muehlmann LA, Ma BC, Longo JPF, Almeida Santos MdFM, Azevedo RB. Aluminum-phthalocyanine chloride associated to poly(methyl vinyl ether-co-maleic anhydride) nanoparticles as a new third-generation photosensitizer for anticancer

- photodynamic therapy. *International journal of nanomedicine*. 2014;9:1199-213. PubMed PMID: 24634582.
18. Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, et al. Photodynamic therapy of cancer: an update. *CA: a cancer journal for clinicians*. 2011 Jul-Aug;61(4):250-81. PubMed PMID: 21617154. Epub 05/26.
 19. Dougherty TJ, Gomer CJ, Henderson BW, Jori G, Kessel D, Korbely M, et al. Photodynamic Therapy. *JNCI: Journal of the National Cancer Institute*. 1998;90(12):889-905.
 20. Dolmans DEJGJ, Fukumura D, Jain RK. Photodynamic therapy for cancer. *Nature Reviews Cancer*. 2003 05/01/online;3:380.
 21. Jayme CC, Calori IR, Cunha EMF, Tedesco AC. Evaluation of aluminum phthalocyanine chloride and DNA interactions for the design of an advanced drug delivery system in photodynamic therapy. *Spectrochimica Acta Part A: Molecular and Biomolecular Spectroscopy*. 2018 2018/08/05;201:242-8.
 22. Dolmans DE, Fukumura D, Jain RK. Photodynamic therapy for cancer. *Nature reviews Cancer*. 2003 May;3(5):380-7. PubMed PMID: 12724736. Epub 2003/05/02. eng.
 23. Amaral LFB, Moriel P, Foglio MA, Mazzola PG. Caryocar brasiliense supercritical CO₂ extract possesses antimicrobial and antioxidant properties useful for personal care products. *BMC complementary and alternative medicine*. 2014;14:73-. PubMed PMID: 24565304.
 24. Sohi KK, Mittal N, Hundal MK, Khanduja KL. Gallic acid, an antioxidant, exhibits antiapoptotic potential in normal human lymphocytes: A Bcl-2 independent mechanism. *Journal of nutritional science and vitaminology*. 2003 Aug;49(4):221-7. PubMed PMID: 14598907. Epub 2003/11/06. eng.
 25. Guimaraes TA, Farias LC, Fraga CA, Feltenberger JD, Melo GA, Coletta RD, et al. Evaluation of the antineoplastic activity of gallic acid in oral squamous cell carcinoma under hypoxic conditions. *Anti-cancer drugs*. 2016;27(5):407-16. PubMed PMID: 00001813-201606000-00004.
 26. Bray F, Ferlay J, Soerjomataram I, Siegel RL, Torre LA, Jemal A. Global cancer statistics 2018: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. *CA: a cancer journal for clinicians*. 2018 Nov;68(6):394-424. PubMed PMID: 30207593.
 27. International Agency for Research on Cancer (IARC) [Internet]. 2018 [cited 18/11/2018].
 28. Fraga CA, de Oliveira MV, de Oliveira ES, Barros LO, Santos FB, Gomez RS, et al. A high HIF-1alpha expression genotype is associated with poor prognosis of upper aerodigestive tract carcinoma patients. *Oral Oncol*. 2012 Feb;48(2):130-5. PubMed PMID: 21945343.
 29. De Paula AM, Souza LR, Farias LC, Correa GT, Fraga CA, Eleuterio NB, et al. Analysis of 724 cases of primary head and neck squamous cell carcinoma (HNSCC) with a focus on young patients and p53 immunolocalization. *Oral oncology*. 2009 Sep;45(9):777-82. PubMed PMID: 19359212. Epub 2009/04/11. eng.
 30. Farias LC, Fraga CA, De Oliveira MV, Silva TF, Marques-Silva L, Moreira PR, et al. Effect of age on the association between p16CDKN2A methylation and DNMT3B polymorphism in head and neck carcinoma and patient survival. *International journal of oncology*. 2010 Jul;37(1):167-76. PubMed PMID: 20514408. Epub 2010/06/02. eng.
 31. Righini CA, Karkas A, Morel N, Soriano E, Reyt E. [Risk factors for cancers of the oral cavity, pharynx (cavity excluded) and larynx]. *Presse medicale (Paris, France : 1983)*. 2008 Sep;37(9):1229-40. PubMed PMID: 18508229. Epub 2008/05/30. Facteurs de risque des cancers de la cavite buccale, du pharynx (cavum exclu) et du larynx. fre.

32. Radoï L, Menvielle G, Cyr D, Lapôtre-Ledoux B, Stücker I, Luce D, et al. Population attributable risks of oral cavity cancer to behavioral and medical risk factors in France: results of a large population-based case–control study, the ICARE study. *BMC Cancer*. 2015 10/31 05/04/received 10/23/accepted;15:827. PubMed PMID: PMC4628276.
33. Min S-K, Choi SW, Ha J, Park JY, Won Y-J, Jung K-W. Conditional relative survival of oral cavity cancer: Based on Korean Central Cancer Registry. *Oral Oncology*. 2017 9//;72:73-9.
34. Guimaraes TA, Farias LC, Santos ES, de Carvalho Fraga CA, Orsini LA, de Freitas Teles L, et al. Metformin increases PDH and suppresses HIF-1alpha under hypoxic conditions and induces cell death in oral squamous cell carcinoma. *Oncotarget*. 2016 Aug 23;7(34):55057-68. PubMed PMID: 27474170. Pubmed Central PMCID: 5342401.
35. Guimaraes TA, Farias LC, Fraga CA, Feltenberger JD, Melo GA, Coletta RD, et al. Evaluation of the antineoplastic activity of gallic acid in oral squamous cell carcinoma under hypoxic conditions. *Anti-cancer drugs*. 2016 Jun;27(5):407-16. PubMed PMID: 26849170.
36. Shah JP, Gil Z. Current concepts in management of oral cancer – Surgery. *Oral Oncology*. 2009 2009/04/01//;45(4):394-401.
37. de Souza MG, de Jesus SF, Santos EM, Gomes ESB, de Paulo Santiago Filho A, Santos EMS, et al. Radiation Therapy Reduced Blood Levels of LDH, HIF-1alpha, and miR-210 in OSCC. *Pathology oncology research : POR*. 2018 Nov 8. PubMed PMID: 30406875.
38. Castilho-Fernandes A, Lopes TG, Primo FL, Pinto MR, Tedesco AC. Photodynamic process induced by chloro-aluminum phthalocyanine nanoemulsion in glioblastoma. *Photodiagnosis and Photodynamic Therapy*. 2017 2017/09/01//;19:221-8.
39. Agostinis P, Berg K, Cengel KA, Foster TH, Girotti AW, Gollnick SO, et al. Photodynamic therapy of cancer: an update. *CA: a cancer journal for clinicians*. 2011 Jul-Aug;61(4):250-81. PubMed PMID: 21617154. Pubmed Central PMCID: PMC3209659. Epub 2011/05/28. eng.
40. Huang Z, Xu H, Meyers AD, Musani AI, Wang L, Tagg R, et al. Photodynamic therapy for treatment of solid tumors--potential and technical challenges. *Technology in cancer research & treatment*. 2008 Aug;7(4):309-20. PubMed PMID: 18642969. Pubmed Central PMCID: Pmc2593637. Epub 2008/07/23. eng.
41. Py-Daniel KR, Namban JS, de Andrade LR, de Souza PEN, Paterno LG, Azevedo RB, et al. Highly efficient photodynamic therapy colloidal system based on chloroaluminum phthalocyanine/pluronic micelles. *European Journal of Pharmaceutics and Biopharmaceutics*. 2016 2016/06/01//;103:23-31.
42. Schmidt MH, Meyer GA, Reichert KW, Cheng J, Krouwer HG, Ozker K, et al. Evaluation of photodynamic therapy near functional brain tissue in patients with recurrent brain tumors. *Journal of neuro-oncology*. 2004 Mar-Apr;67(1-2):201-7. PubMed PMID: 15072468. Epub 2004/04/10. eng.
43. Quirk BJ, Brandal G, Donlon S, Vera JC, Mang TS, Foy AB, et al. Photodynamic therapy (PDT) for malignant brain tumors – Where do we stand? *Photodiagnosis and Photodynamic Therapy*. 2015 2015/09/01//;12(3):530-44.
44. Foote CS. Definition of type I and type II photosensitized oxidation. *Photochemistry and photobiology*. 1991 Nov;54(5):659. PubMed PMID: 1798741. Epub 1991/11/01. eng.
45. Nunes SM, Sguilla FS, Tedesco AC. Photophysical studies of zinc phthalocyanine and chloroaluminum phthalocyanine incorporated into liposomes in the presence of additives. *Brazilian journal of medical and biological research = Revista brasileira de pesquisas medicas e biologicas*. 2004 Feb;37(2):273-84. PubMed PMID: 14762584. Epub 2004/02/06. eng.
46. Muehlmann LA, Ma BC, Longo JP, Almeida Santos Mde F, Azevedo RB. Aluminum-phthalocyanine chloride associated to poly(methyl vinyl ether-co-maleic anhydride)

- nanoparticles as a new third-generation photosensitizer for anticancer photodynamic therapy. *International journal of nanomedicine*. 2014;9:1199-213. PubMed PMID: 24634582. Pubmed Central PMCID: PMC3952896. Epub 2014/03/19. eng.
47. Muehlmann LA, Rodrigues MC, Longo JP, Garcia MP, Py-Daniel KR, Veloso AB, et al. Aluminium-phthalocyanine chloride nanoemulsions for anticancer photodynamic therapy: Development and in vitro activity against monolayers and spheroids of human mammary adenocarcinoma MCF-7 cells. *Journal of nanobiotechnology*. 2015 May 13;13:36. PubMed PMID: 25966866. Pubmed Central PMCID: PMC4455699. Epub 2015/05/15. eng.
48. Morgado LF, Travolo ARF, Muehlmann LA, Narcizo PS, Nunes RB, Pereira PAG, et al. Photodynamic Therapy treatment of onychomycosis with Aluminium-Phthalocyanine Chloride nanoemulsions: A proof of concept clinical trial. *Journal of photochemistry and photobiology B, Biology*. 2017 Aug;173:266-70. PubMed PMID: 28622558.
49. Chan SW, Lim CJ, Guo K, Ng CP, Lee I, Hunziker W, et al. A role for TAZ in migration, invasion, and tumorigenesis of breast cancer cells. *Cancer research*. 2008 Apr 15;68(8):2592-8. PubMed PMID: 18413727. Epub 2008/04/17. eng.
50. Kasibhatla S, Amarante-Mendes GP, Finucane D, Brunner T, Bossy-Wetzel E, Green DR. Acridine Orange/Ethidium Bromide (AO/EB) Staining to Detect Apoptosis. *Cold Spring Harbor Protocols*. 2006 August 1, 2006;2006(3):pdb.prot4493.
51. Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. *Nature methods*. 2012 Jul;9(7):671-5. PubMed PMID: 22930834. Pubmed Central PMCID: PMC5554542. Epub 2012/08/30. eng.
52. Castano AP, Demidova TN, Hamblin MR. Mechanisms in photodynamic therapy: part one—photosensitizers, photochemistry and cellular localization. *Photodiagnosis and photodynamic therapy*. 2004;1(4):279-93. PubMed PMID: PMC4108220.
53. Fonseca-Silva T, Farias LC, Cardoso CM, Souza LR, Carvalho Fraga CA, Oliveira MV, et al. Analysis of p16(CDKN2A) methylation and HPV-16 infection in oral mucosal dysplasia. *Pathobiology : journal of immunopathology, molecular and cellular biology*. 2012;79(2):94-100. PubMed PMID: 22285991. Epub 2012/01/31. eng.
54. Pereira CS, Oliveira MV, Fraga CA, Barros LO, Domingos PL, Roy A, et al. Impact of the epithelial dysplasia grading and Ki67 proliferation index in the adjacent non-malignant mucosa on recurrence and survival in head and neck squamous cell carcinoma. *Pathology, research and practice*. 2012 Nov 15;208(11):651-6. PubMed PMID: 22995634. Epub 2012/09/22. eng.
55. de Oliveira MV, Pereira Gomes EP, Pereira CS, de Souza LR, Barros LO, Mendes DC, et al. Prognostic value of microvessel density and p53 expression on the locoregional metastasis and survival of the patients with head and neck squamous cell carcinoma. *Applied immunohistochemistry & molecular morphology : AIMM*. 2013 Oct;21(5):444-51. PubMed PMID: 23343952.
56. Barclay CW, Foster EC, Taylor CL. Restorative aspects of oral cancer reconstruction. *British dental journal*. 2018 Nov 9;225(9):848-54. PubMed PMID: 30412540.
57. Justus CR, Leffler N, Ruiz-Echevarria M, Yang LV. In vitro cell migration and invasion assays. *Journal of visualized experiments : JoVE*. 2014 (88):51046. PubMed PMID: 24962652.
58. Domingos PLB, Souza MG, Guimaraes TA, Santos ES, Farias LC, de Carvalho Fraga CA, et al. Hypoxia reduces the E-cadherin expression and increases OSCC cell migration regardless of the E-cadherin methylation profile. *Pathology, research and practice*. 2017 May;213(5):496-501. PubMed PMID: 28285966.
59. Jabir MS, Taha AA, Sahib UI, Taqi ZJ, Al-Shammari AM, Salman AS. Novel of nano delivery system for Linalool loaded on gold nanoparticles conjugated with CALNN peptide for application in drug uptake and induction of cell death on breast cancer cell line. *Materials*

- science & engineering C, *Materials for biological applications*. 2019 Jan 1;94:949-64. PubMed PMID: 30423784. Epub 2018/11/15. eng.
60. Debele TA, Peng S, Tsai H-C. Drug Carrier for Photodynamic Cancer Therapy. *International journal of molecular sciences*. 2015;16(9):22094-136. PubMed PMID: 26389879.
61. Sobocki M, Mrouj K, Camasses A, Parisi N, Nicolas E, Llères D, et al. The cell proliferation antigen Ki-67 organises heterochromatin. *eLife*. 2016 03/07 12/21/received 03/06/accepted;5:e13722. PubMed PMID: PMC4841783.
62. Scholzen T, Gerdes J. The Ki-67 protein: from the known and the unknown. *Journal of cellular physiology*. 2000 Mar;182(3):311-22. PubMed PMID: 10653597. Epub 2000/02/01. eng.
63. Jing Y, Yang Y, Hao F, Song Y, Zhang X, Zhang Y, et al. Higher Ki67 expression in fibroblast like cells at invasive front indicates better clinical outcomes in oral squamous cell carcinoma patients. *Bioscience reports*. 2018 Dec 21;38(6). PubMed PMID: 30341240. Epub 2018/10/21. eng.
64. Hasegawa T, Yamamoto S, Yokoyama R, Umeda T, Matsuno Y, Hirohashi S. Prognostic significance of grading and staging systems using MIB-1 score in adult patients with soft tissue sarcoma of the extremities and trunk. *Cancer*. 2002 Aug 15;95(4):843-51. PubMed PMID: 12209729. Epub 2002/09/05. eng.
65. Kausch I, Lingnau A, Endl E, Sellmann K, Deinert I, Ratliff TL, et al. Antisense treatment against Ki-67 mRNA inhibits proliferation and tumor growth in vitro and in vivo. *International journal of cancer*. 2003 Jul 10;105(5):710-6. PubMed PMID: 12740923. Epub 2003/05/13. eng.
66. Li C-F, Chen L-T, Lan J, Chou F-F, Lin C-Y, Chen Y-Y, et al. AMACR amplification and overexpression in primary imatinib-naïve gastrointestinal stromal tumors: a driver of cell proliferation indicating adverse prognosis. *Oncotarget*. 2014 10/18 08/13/received 10/18/accepted;5(22):11588-603. PubMed PMID: PMC4294386.
67. Mohamed AA, Abbas MY, Alharbi H, Babiker AY. Assessment of Expression of Ki-67 in Benign and Malignant Prostatic Lesions among Sudanese Patients. *Open Access Macedonian Journal of Medical Sciences*. 2018;6(10):1809-12. PubMed PMID: PMC6236043.
68. Zhu J, Sammons MA, Donahue G, Dou Z, Vedadi M, Getlik M, et al. Prevalent p53 mutants co-opt chromatin pathways to drive cancer growth. *Nature*. 2015 09/02;525(7568):206-11. PubMed PMID: PMC4568559.
69. Ozaki T, Nakagawara A. Role of p53 in Cell Death and Human Cancers. *Cancers*. 2011;3(1):994-1013. PubMed PMID: 24212651.
70. Pinheiro UB, Fraga CA, Mendes DC, Farias LC, Cardoso CM, Silveira CM, et al. Fuzzy clustering demonstrates that codon 72 SNP rs1042522 of TP53 gene associated with HNSCC but not with prognoses. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2015 Dec;36(12):9259-65. PubMed PMID: 26099726.
71. Souza LR, Fonseca-Silva T, Pereira CS, Santos EP, Lima LC, Carvalho HA, et al. Immunohistochemical analysis of p53, APE1, hMSH2 and ERCC1 proteins in actinic cheilitis and lip squamous cell carcinoma. *Histopathology*. 2011 Feb;58(3):352-60. PubMed PMID: 21323960.
72. Pereira T, Brito JAR, Guimaraes ALS, Gomes CC, de Lacerda JCT, de Castro WH, et al. MicroRNA profiling reveals dysregulated microRNAs and their target gene regulatory networks in cemento-ossifying fibroma. *Journal of oral pathology & medicine : official*

- publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology. 2018 Jan;47(1):78-85. PubMed PMID: 29032608.
73. Hientz K, Mohr A, Bhakta-Guha D, Efferth T. The role of p53 in cancer drug resistance and targeted chemotherapy. *Oncotarget*. 2017 11/19 07/10/received 10/13/accepted;8(5):8921-46. PubMed PMID: PMC5352454.
74. Chan JY-H, Chen Y-C, Liu S-T, Chou W-Y, Ho C-L, Huang S-M. Characterization of a new mouse p53 variant: loss-of-function and gain-of-function. *Journal of biomedical science*. 2014;21(1):40-. PubMed PMID: 24884657.
75. Shield KD, Ferlay J, Jemal A, Sankaranarayanan R, Chaturvedi AK, Bray F, et al. The global incidence of lip, oral cavity, and pharyngeal cancers by subsite in 2012. *CA: a cancer journal for clinicians*. 2017 Jan;67(1):51-64. PubMed PMID: 28076666.
76. Souza RL, Fonseca-Fonseca T, Oliveira-Santos CC, Correa GT, Santos FB, Cardoso CM, et al. Lip squamous cell carcinoma in a Brazilian population: epidemiological study and clinicopathological associations. *Med Oral Patol Oral Cir Bucal*. 2011 Sep 1;16(6):e757-62. PubMed PMID: 21196879. Epub 2011/01/05. eng.
77. Marques-Silva L, Farias LC, Fraga CA, de Oliveira MV, Cardos CM, Fonseca-Silva T, et al. HPV-16/18 detection does not affect the prognosis of head and neck squamous cell carcinoma in younger and older patients. *Oncology letters*. 2012 Apr 1;3(4):945-9. PubMed PMID: 22741024. Pubmed Central PMCID: Pmc3362356. Epub 2012/06/29. eng.
78. Correa GT, Bandeira GA, Cavalcanti BG, de Carvalho Fraga CA, dos Santos EP, Silva TF, et al. Association of -308 TNF-alpha promoter polymorphism with clinical aggressiveness in patients with head and neck squamous cell carcinoma. *Oral oncology*. 2011 Sep;47(9):888-94. PubMed PMID: 21788151. Epub 2011/07/27. eng.
79. de Carvalho Fraga CA, Farias LC, de Oliveira MV, Domingos PL, Pereira CS, Silva TF, et al. Increased VEGFR2 and MMP9 protein levels are associated with epithelial dysplasia grading. *Pathology, research and practice*. 2014 Dec;210(12):959-64. PubMed PMID: 25441661. Epub 2014/12/03. eng.
80. Pinheiro UB, de Carvalho Fraga CA, Mendes DC, Marques-Silva L, Farias LC, de Souza MG, et al. p16 (CDKN2A) SNP rs11515 was not associated with head and neck carcinoma. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2014 Jun;35(6):6113-8. PubMed PMID: 24633888. Epub 2014/03/19. eng.
81. de Carvalho Fraga CA, Farias LC, Jones KM, Batista de Paula AM, Guimaraes ALS. Angiotensin-Converting Enzymes (ACE and ACE2) as Potential Targets for Malignant Epithelial Neoplasia: Review and Bioinformatics Analyses Focused in Oral Squamous Cell Carcinoma. *Protein and peptide letters*. 2017 Nov 17;24(9):784-92. PubMed PMID: 28814250. Epub 2017/08/02. eng.
82. van der Waal I. Oral potentially malignant disorders: is malignant transformation predictable and preventable? *Med Oral Patol Oral Cir Bucal*. 2014 Jul 1;19(4):e386-90. PubMed PMID: 24905952. Pubmed Central PMCID: 4119315.
83. Vairaktaris E, Spyridonidou S, Papakosta V, Vylliotis A, Lazaris A, Perrea D, et al. The hamster model of sequential oral oncogenesis. *Oral oncology*. 2008 Apr;44(4):315-24. PubMed PMID: 18061531. Epub 2007/12/07. eng.
84. Liu X, He R, Chen W. [A rat model of tongue mucosa squamous cell carcinoma induced by oral administration of 4NQO in drinking water]. *Zhonghua kou qiang yi xue za zhi = Zhonghua kouqiang yixue zazhi = Chinese journal of stomatology*. 1999 Nov;34(6):354-6. PubMed PMID: 11776878. Epub 2002/01/05. chi.

85. Dayan D, Hirshberg A, Kaplan I, Rotem N, Bodner L. Experimental tongue cancer in desalivated rats. *Oral oncology*. 1997 Mar;33(2):105-9. PubMed PMID: 9231167. Epub 1997/03/01. eng.
86. Vitale-Cross L, Czerninski R, Amornphimoltham P, Patel V, Molinolo AA, Gutkind JS. Chemical carcinogenesis models for evaluating molecular-targeted prevention and treatment of oral cancer. *Cancer prevention research (Philadelphia, Pa)*. 2009 May;2(5):419-22. PubMed PMID: 19401522. Epub 2009/04/30. eng.
87. Vitale-Cross L, Molinolo AA, Martin D, Younis RH, Maruyama T, Patel V, et al. Metformin prevents the development of oral squamous cell carcinomas from carcinogen-induced premalignant lesions. *Cancer prevention research (Philadelphia, Pa)*. 2012 Apr;5(4):562-73. PubMed PMID: 22467081. Pubmed Central PMCID: PMC3429367. Epub 2012/04/03. eng.
88. Nagler R, Dayan D. The dual role of saliva in oral carcinogenesis. *Oncology*. 2006;71(1-2):10-7. PubMed PMID: 17344667. Epub 2007/03/09. eng.
89. Barcessat AR, Huang I, Rabelo GD, Rosin FC, Ferreira LG, de Cerqueira Luz JG, et al. Systemic toxic effects during early phases of topical 4-NQO-induced oral carcinogenesis in rats. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2014 Nov;43(10):770-7. PubMed PMID: 24931357.
90. Mendenhall WM, Dagan R, Bryant CM, Fernandes RP. Radiation Oncology for Head and Neck Cancer: Current Standards and Future Changes. *Oral and maxillofacial surgery clinics of North America*. 2019 Feb;31(1):31-8. PubMed PMID: 30454789.
91. Algazi AP, Grandis JR. Head and neck cancer in 2016: A watershed year for improvements in treatment? *Nature reviews Clinical oncology*. 2017 Feb;14(2):76-8. PubMed PMID: 27922045. Epub 2016/12/07. eng.
92. Lasrado S, Moras K, Pinto GJ, Bhat M, Hegde S, Sathian B, et al. Role of concomitant chemoradiation in locally advanced head and neck cancers. *Asian Pacific journal of cancer prevention : APJCP*. 2014;15(10):4147-52. PubMed PMID: 24935361. Epub 2014/06/18. eng.
93. Hiraishi Y, Wada T, Nakatani K, Tojyo I, Matsumoto T, Kiga N, et al. EGFR inhibitor enhances cisplatin sensitivity of oral squamous cell carcinoma cell lines. *Pathology oncology research : POR*. 2008;14(1):39-43. PubMed PMID: 18347929. Epub 03/07.
94. Bhuvanewari R, Ng QF, Thong PS, Soo KC. Nimotuzumab increases the anti-tumor effect of photodynamic therapy in an oral tumor model. *Oncotarget*. 2015 May 30;6(15):13487-505. PubMed PMID: 25918252. Pubmed Central PMCID: PMC4537029. Epub 2015/04/29. eng.
95. Sankaranarayanan R, Mathew B, Jacob BJ, Thomas G, Somanathan T, Pisani P, et al. Early findings from a community-based, cluster-randomized, controlled oral cancer screening trial in Kerala, India. The Trivandrum Oral Cancer Screening Study Group. *Cancer*. 2000 Feb 1;88(3):664-73. PubMed PMID: 10649262.
96. Frenandez Garrote L, Sankaranarayanan R, Lence Anta JJ, Rodriguez Salva A, Maxwell Parkin D. An evaluation of the oral cancer control program in Cuba. *Epidemiology*. 1995 Jul;6(4):428-31. PubMed PMID: 7548355.
97. Chuang SL, Su WW, Chen SL, Yen AM, Wang CP, Fann JC, et al. Population-based screening program for reducing oral cancer mortality in 2,334,299 Taiwanese cigarette smokers and/or betel quid chewers. *Cancer*. 2017 May 1;123(9):1597-609. PubMed PMID: 28055109.
98. Chang SL, Lo CH, Peng HL, Chen CR, Wu SC, Chen SC. Factors associated with continued smoking after treatment of oral cavity cancer: An age and survival time-matched study. *Journal of advanced nursing*. 2018 Apr;74(4):926-34. PubMed PMID: 29148210.


99. Hamadah O, Hepburn S, Thomson PJ. Effects of active non-smoking programmes on smoking behaviour in oral precancer patients. *International journal of oral and maxillofacial surgery*. 2007 Aug;36(8):706-11. PubMed PMID: 17448634.
100. Yamamoto K, Kitayama W, Denda A, Morisaki A, Kuniyasu H, Kirita T. Inhibitory effects of selective cyclooxygenase-2 inhibitors, nimesulide and etodolac, on the development of squamous cell dysplasias and carcinomas of the tongue in rats initiated with 4-nitroquinoline 1-oxide. *Cancer letters*. 2003 Sep 25;199(2):121-9. PubMed PMID: 12969784. Epub 2003/09/13. eng.
101. Donati M, Conforti A, Lenti MC, Capuano A, Bortolami O, Motola D, et al. Risk of acute and serious liver injury associated to nimesulide and other NSAIDs: data from drug-induced liver injury case-control study in Italy. *British journal of clinical pharmacology*. 2016 Jul;82(1):238-48. PubMed PMID: 26991794. Pubmed Central PMCID: 4917796.
102. Cabrera E, Levenson J, Armentano R, Barra J, Pichel R, Simon AC. Aortic pulsatile pressure and diameter response to intravenous perfusions of angiotensin, norepinephrine, and epinephrine in conscious dogs. *Journal of cardiovascular pharmacology*. 1988 Dec;12(6):643-9. PubMed PMID: 2467081.
103. Lalau JD. Lactic acidosis induced by metformin: incidence, management and prevention. *Drug safety*. 2010 Sep 1;33(9):727-40. PubMed PMID: 20701406.
104. Subramanian AP, Jaganathan SK, Mandal M, Supriyanto E, Muhamad II. Gallic acid induced apoptotic events in HCT-15 colon cancer cells. *World journal of gastroenterology*. 2016;22(15):3952-61. PubMed PMID: 27099438. Epub 04/21.
105. Daglia M, Di Lorenzo A, Nabavi SF, Talas ZS, Nabavi SM. Polyphenols: well beyond the antioxidant capacity: gallic acid and related compounds as neuroprotective agents: you are what you eat! *Current pharmaceutical biotechnology*. 2014;15(4):362-72. PubMed PMID: 24938889. Epub 2014/06/19. eng.
106. de Oliveira TS, Thomaz DV, da Silva Neri HF, Cerqueira LB, Garcia LF, Gil HPV, et al. Neuroprotective Effect of Caryocar brasiliense Camb. Leaves Is Associated with Anticholinesterase and Antioxidant Properties. *Oxidative medicine and cellular longevity*. 2018;2018:9842908-. PubMed PMID: 30420910.
107. Verma S, Singh A, Mishra A. Gallic acid: molecular rival of cancer. *Environmental toxicology and pharmacology*. 2013 May;35(3):473-85. PubMed PMID: 23501608. Epub 2013/03/19. eng.
108. Santos EMS, da Rocha RG, Santos HO, Guimaraes TA, de Carvalho Fraga CA, da Silveira LH, et al. Gallic acid modulates phenotypic behavior and gene expression in oral squamous cell carcinoma cells by interfering with leptin pathway. *Pathology, research and practice*. 2018 Jan;214(1):30-7. PubMed PMID: 29254802. Epub 2017/12/20. eng.
109. Lu Y, Jiang F, Jiang H, Wu K, Zheng X, Cai Y, et al. Gallic acid suppresses cell viability, proliferation, invasion and angiogenesis in human glioma cells. *European journal of pharmacology*. 2010 Sep 1;641(2-3):102-7. PubMed PMID: 20553913. Pubmed Central PMCID: PMC3003697. Epub 2010/06/18. eng.
110. Ho HH, Chang CS, Ho WC, Liao SY, Lin WL, Wang CJ. Gallic acid inhibits gastric cancer cells metastasis and invasive growth via increased expression of RhoB, downregulation of AKT/small GTPase signals and inhibition of NF-kappaB activity. *Toxicology and applied pharmacology*. 2013 Jan 1;266(1):76-85. PubMed PMID: 23153558. Epub 2012/11/17. eng.
111. Liu Z, Schwimer J, Liu D, Lewis J, Greenway FL, York DA, et al. Gallic acid is partially responsible for the antiangiogenic activities of Rubus leaf extract. *Phytotherapy research : PTR*. 2006 Sep;20(9):806-13. PubMed PMID: 16835875. Epub 2006/07/13. eng.

112. Inoue M, Suzuki R, Sakaguchi N, Li Z, Takeda T, Ogihara Y, et al. Selective induction of cell death in cancer cells by gallic acid. *Biological & pharmaceutical bulletin*. 1995 Nov;18(11):1526-30. PubMed PMID: 8593472. Epub 1995/11/01. eng.
113. Sobrinho Santos EM, Guimaraes TA, Santos HO, Cangussu LMB, de Jesus SF, Fraga CAC, et al. Leptin acts on neoplastic behavior and expression levels of genes related to hypoxia, angiogenesis, and invasiveness in oral squamous cell carcinoma. *Tumour biology : the journal of the International Society for Oncodevelopmental Biology and Medicine*. 2017 May;39(5):1010428317699130. PubMed PMID: 28459203. Epub 2017/05/02. eng.
114. Karimi-Khouzani O, Heidarian E, Amini SA. Anti-inflammatory and ameliorative effects of gallic acid on fluoxetine-induced oxidative stress and liver damage in rats. *Pharmacological Reports*. 2017 2017/08/01;69(4):830-5.
115. Han ED, MacFarlane RC, Mulligan AN, Scafidi J, Davis AE, 3rd. Increased vascular permeability in C1 inhibitor-deficient mice mediated by the bradykinin type 2 receptor. *The Journal of clinical investigation*. 2002 Apr;109(8):1057-63. PubMed PMID: 11956243. Pubmed Central PMCID: 150945.
116. Mendes-Junior LG, Freitas-Lima LC, Oliveira JR, Melo MB, Feltenberger JD, Brandi IV, et al. The usefulness of short-term high-fat/high salt diet as a model of metabolic syndrome in mice. *Lasers in medical science*. 2018 Sep 15;209:341-8. PubMed PMID: 30118771. Epub 2018/06/28. eng.
117. Gomes EP, Aguiar JC, Fonseca-Silva T, Dias LC, Moura-Boas KP, Roy A, et al. Diazepam reverses the alveolar bone loss and hippocampal interleukin-1beta and interleukin-6 enhanced by conditioned fear stress in ligature-induced periodontal disease in rats. *Journal of periodontal research*. 2013 Apr;48(2):151-8. PubMed PMID: 22891744. Epub 2012/08/16. eng.
118. Schneider CA, Rasband WS, Eliceiri KW. NIH Image to ImageJ: 25 years of image analysis. *Nature Methods*. 2012 06/28/online;9:671.
119. Warnakulasuriya S, Reibel J, Bouquot J, Dabelsteen E. Oral epithelial dysplasia classification systems: predictive value, utility, weaknesses and scope for improvement. *Journal of oral pathology & medicine : official publication of the International Association of Oral Pathologists and the American Academy of Oral Pathology*. 2008 Mar;37(3):127-33. PubMed PMID: 18251935.
120. Li C-C, Shen Z, Bavarian R, Yang F, Bhattacharya A. Oral Cancer: Genetics and the Role of Precision Medicine. *Dental Clinics of North America*. 2018 2018/01/01;62(1):29-46.
121. Correa GT, Bandeira GA, Cavalcanti BG, Santos FB, Rodrigues Neto JF, Guimaraes AL, et al. Analysis of ECOG performance status in head and neck squamous cell carcinoma patients: association with sociodemographical and clinical factors, and overall survival. *Supportive care in cancer : official journal of the Multinational Association of Supportive Care in Cancer*. 2012 Nov;20(11):2679-85. PubMed PMID: 22314971. Epub 2012/02/09. eng.
122. Doan KV, Ko CM, Kinyua AW, Yang DJ, Choi YH, Oh IY, et al. Gallic acid regulates body weight and glucose homeostasis through AMPK activation. *Endocrinology*. 2015 Jan;156(1):157-68. PubMed PMID: 25356824.
123. Lucena SR, Salazar N, Gracia-Cazaña T, Zamarrón A, González S, Juarranz Á, et al. Combined Treatments with Photodynamic Therapy for Non-Melanoma Skin Cancer. *International journal of molecular sciences*. 2015;16(10):25912-33. PubMed PMID: 26516853.
124. Chao J, Huo TI, Cheng HY, Tsai JC, Liao JW, Lee MS, et al. Gallic acid ameliorated impaired glucose and lipid homeostasis in high fat diet-induced NAFLD mice. *PloS one*. 2014;9(2):e96969. PubMed PMID: 24918580. Pubmed Central PMCID: 4053315.

125. Li D, Liu Z, Zhao W, Xi Y, Niu F. A straightforward method to determine the cytotoxic and cytopathic effects of the functional groups of gallic acid. *Process Biochemistry*. 2011 2011/11/01/;46(11):2210-4.
126. Sun G, Zhang S, Xie Y, Zhang Z, Zhao W. Gallic acid as a selective anticancer agent that induces apoptosis in SMMC-7721 human hepatocellular carcinoma cells. *Oncology Letters*. 2016 10/30 10/28/received 08/20/accepted;11(1):150-8. PubMed PMID: PMC4727056.
127. Inoue M, Suzuki R, Koide T, Sakaguchi N, Ogihara Y, Yabu Y. Antioxidant, Gallic Acid, Induces Apoptosis in HL-60RG Cells. *Biochemical and Biophysical Research Communications*. 1994 1994/10/31/;204(2):898-904.
128. Weng Y-P, Hung P-F, Ku W-Y, Chang C-Y, Wu B-H, Wu M-H, et al. The inhibitory activity of gallic acid against DNA methylation: application of gallic acid on epigenetic therapy of human cancers. *Oncotarget*. 2018 12/07 07/11/received 11/13/accepted;9(1):361-74. PubMed PMID: PMC5787471.
129. Rattanata N, Daduang S, Wongwattanakul M, Leelayuwat C, Limpaboon T, Lekphrom R, et al. Gold Nanoparticles Enhance the Anticancer Activity of Gallic Acid against Cholangiocarcinoma Cell Lines. *Asian Pacific journal of cancer prevention : APJCP*. 2015;16(16):7143-7. PubMed PMID: 26514503. Epub 2015/10/31. eng.
130. Ho H-H, Chang C-S, Ho W-C, Liao S-Y, Lin W-L, Wang C-J. Gallic acid inhibits gastric cancer cells metastasis and invasive growth via increased expression of RhoB, downregulation of AKT/small GTPase signals and inhibition of NF- κ B activity. *Toxicology and applied pharmacology*. 2013 2013/01/01/;266(1):76-85.
131. You BR, Park WH. Gallic acid-induced lung cancer cell death is related to glutathione depletion as well as reactive oxygen species increase. *Toxicology in Vitro*. 2010 2010/08/01/;24(5):1356-62.
132. de Paiva Gonçalves V, Ortega AAC, Guimarães MR, Curylofo FA, Junior CR, Ribeiro DA, et al. Chemopreventive Activity of Systemically Administered Curcumin on Oral Cancer in the 4-Nitroquinoline 1-Oxide Model. *Journal of Cellular Biochemistry*. 2015;116(5):787-96.
133. Hilbig J, Policarpi PdB, Grinevicius VMAdS, Mota NSRS, Toaldo IM, Luiz MTB, et al. Aqueous extract from pecan nut [*Carya illinoensis* (Wangenh) C. Koch] shell show activity against breast cancer cell line MCF-7 and Ehrlich ascites tumor in Balb-C mice. *Journal of Ethnopharmacology*. 2018 2018/01/30/;211:256-66.
134. de Oliveira LS, Thome GR, Lopes TF, Reichert KP, de Oliveira JS, da Silva Pereira A, et al. Effects of gallic acid on delta - aminolevulinic dehydratase activity and in the biochemical, histological and oxidative stress parameters in the liver and kidney of diabetic rats. *Biomedicine & pharmacotherapy = Biomedecine & pharmacotherapie*. 2016 Dec;84:1291-9. PubMed PMID: 27810786. Epub 2016/11/05. eng.

ANEXOS



UNIVERSIDADE ESTADUAL DE MONTES CLAROS
COMISSÃO DE ÉTICA EM EXPERIMENTAÇÃO E BEM-ESTAR ANIMAL  **Unimontes**

PARECER CONSUBSTANCIADO

Montes Claros, 19 de fevereiro de 2016.

Processo N.º 098

Título do Projeto: Avaliação da atividade antineoplásica de compostos metabólicos secundários em carcinoma de escamosas de boca: uma abordagem *in vivo*

Pesquisador responsável: Dr. André Luiz Sena Guimarães

Histórico

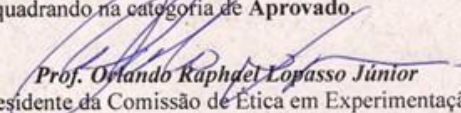
O carcinoma de células escamosas de boca (CCEB) é a neoplasia maligna mais frequente da região de cabeça e pescoço, representa um dos principais problemas de saúde pública. No Brasil é o sexto tipo de câncer mais prevalente, mas também em muitos países do mundo devido a sua alta mortalidade e constante mutilação. Evidências demonstraram que diversas alterações genéticas, epigenéticas e metabólicas estão associadas à patogênese desta neoplasia. Como consequência, o organismo perde o controle da proliferação celular sobre as células do CCEB. Como característica importante possui áreas hipóxicas, o que dificulta o efeito da radioterapia. Por isso, atualmente a combinação entre quimioterapia e radioterapia têm se mostrado necessário para o tratamento do CCEB. Porém, provoca mais efeitos colaterais nos pacientes com CCEB e apresenta custo elevado. Assim, faz-se necessário o desenvolvimento de pesquisas que busquem o desenvolvimento de novas estratégias terapêuticas. Estudos recentes demonstraram que o ácido gálico (AG) têm se mostrado agente terapêutico em potencial para o CCEB, uma vez que, apresenta propriedades antiproliferativa e citotóxica em vários tipos células neoplásicas

Mérito

O projeto justifica-se, por buscar uma nova abordagem de tratamento com o objetivo de amenizar o comportamento invasivo da doença. Objetivando avaliar a atividade antineoplásica dos metabólitos secundários: Ácido gálico, Metformina, Ácido Arjunólico (AA) e Ácido Betulínico (AB) no CCEB em modelos *in vivo*.

Parecer

A Comissão de Ética em Experimentação e Bem-Estar Animal da Unimontes analisou o processo 098 e entende que o protocolo de procedimentos preenche todos os requisitos éticos do CEEBEA/Unimontes enquadrando na categoria de **Aprovado**.


Prof. Orlando Raphael Lopes Júnior
Presidente da Comissão de Ética em Experimentação
e Bem-Estar Animal da UNIMONTES